Evidence-based research in pediatric nutrition: focus on the supplementation of polyunsaturated fatty acids and prebiotics

Doctoral (PhD) Dissertation

Szimonetta Lohner M.D.

Programme leader: Prof. Dr. Tamás Decsi

Theme leader: Prof. Dr. Tamás Decsi

University of Pécs, Medical School Pécs, Hungary

Pécs, 2014

Abbreviations

LCPUFA long-chain polyunsaturated fatty acid

LA linoleic acid (C18:2n-6)

GLA γ-linolenic acid (C18:3n-6)

DHGLA dihomo-γ-linolenic acid (C20:3n-6)

AA arachidonic acid (C20:4n-6)

ALA α -linolenic acid (C18:3n-3)

EPA eicosapentaenoic acid (C20:5n-3)

DPA docosapentaenoic acid (C22:5n-3)

DHA docosahexaenoic acid (C22:6n-3)

PL phospholipid

TG triacylglycerol

PC phosphatidylcholine

CE cholesteryl ester

PEA phospatidylethanolamine

FFA free fatty acids

MD mean difference

CI confidence interval

RCT randomized controlled trial

RR rate ratio

PKU phenylketonuria

Phe phenylalanine

HMO human milk oligosaccharides

GOS galacto-oligosaccharides

FOS fructo-oligosaccharides

RTI acute respiratory tract infections

URTI upper respiratory tract infections

LRTI lower respiratory tract infections

GITI gastrointestinal infections

UTI urinary tract infections

Table of contents

1. Introduction	Ь
2. Methodical aspects of conducting a systematic review and meta-analysis	10
2.1 The PRISMA Statement	10
2.2 Framing the question and deciding on important outcomes	11
2.3. Searching for studies, selecting studies and collecting data	11
2.4 Assessment of the validity of the findings	12
2.5 Data synthesis and meta-analysis	12
2.6 Presenting results	14
3. Aims	16
4. Meta-analyses	17
4.1. Gender differences in long-chain polyunsaturated fatty acid statu	ıs of
healthy subjects: a systematic review and meta-analysis	17
4.1.1. Inclusion criteria	17
4.1.2. Search strategy	17
4.1.3. Data extraction	20
4.1.4. Statistical analysis	20
4.1.5. Results	21
4.1.6. Discussion	35
4.2. Long-chain polyunsaturated fatty acid status in patients with phen	ylke-
tonuria as compared to healthy controls: a systematic review and meta-analysis	38
4.2.1. Inclusion criteria	38
4.2.2. Search strategy	38
4.2.3. Data extraction	39
4.2.4. Statistical analysis	40
4.2.5. Results	41
4.2.6. Discussion	47

4.3 Prebiotics in healthy infants and children for prevention of	f acute infec
tious diseases: a systematic review and meta-analysis	49
4.3.1. Inclusion criteria	49
4.3.2. Search Strategy	49
4.3.3. Data extraction and management	52
4.3.4 Statistical analysis	52
4.3.5. Results	53
4.3.6. Discussion	62
5. Novel findings and practical applications	65
6. Acknowledgement	66
7. List of references	67

1. Introduction

Evidence-based research in pediatric nutrition

Evidence-based medicine is defined as the conscientious, explicit, and reflective use of current best evidence in making decisions about the care of individual patients. It involves integrating clinical expertise with the best available external evidence from systematic research, and incorporating this into clinical decision-making (Moyer A et al. 2004).

The number of systematic reviews is increasing rapidly, also in the field of nutrition and pediatric nutrition. In nutrition, systematic reviews has been used or were considered for the formulation of dietary guidelines, the establishment of nutrient reference intakes, the formulation of clinical practice guidelines and community practice guidelines as well as for the evaluation of applications for food and supplement label health claims and for the identification of research needs and priorities (Lichtenstein et al. 2008). Systematic reviews represent a palpable support for evidence-based practitioners to weigh the strength of the evidence and, accordingly, the degree of uncertainly. However, it has to be considered that systematic reviews provide answer to a specific question, which may be only one among many needed to address an important and actual topic. In the field of nutrition, systematic reviews are essential for accurate summarizing of the evidence in the following issues (Szajewska 2013):

- baseline exposure to nutrients (either from food or from supplement intake)
- nutrient status of a population
- bioequivalence of different chemical forms of nutrients
- bioavailability of nutrients and their different chemical forms
- biological functions of a nutrient
- assessing dose-response relationships.

The nutritional role of long-chain polyunsaturated fatty acids

Long-chain-polyunsaturated fatty acids (LCPUFA) have important functions in cell membranes as indispensable building stones for human development and optimal health. Docosahexaenoic acid (DHA) and arachidonic acid (AA) are considered to be the most important functional LCPUFA. They can be provided directly from the diet or can be synthetized from their essential fatty acid precursors alpha-linolenic acid (ALA) and linoleic acid (LA).

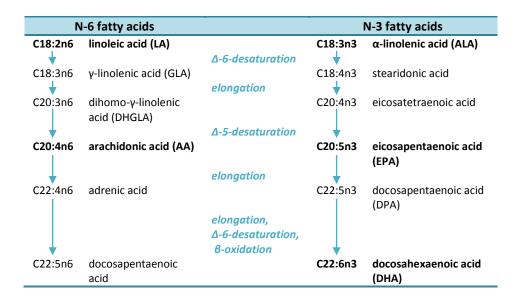


Figure 1. Metabolism of the n-6 and n-3 fatty acids

Among other enzymes, delta-6-desaturase and delta-5-desaturase are required for the formation of the longer-chain metabolites of both n-3 and n-6 series. The competitive desaturation of the n-3 and n-6 series of fatty acids by delta-6-desaturase is of major significance, because this step is considered to be the rate-limiting step of the pathway. The activity of this enzyme is modulated by hormones and by interactions of substrates and metabolic products (Burdge and Wootton 2002; Harnack et al. 2009). The fatty acid composition of cell membrane phospholipids is determined by the fatty acid composition of the diet; both, sufficient intake and the optimal n-3/n-6 fatty acid content of diet are of large importance.

Diseases, (such as diabetes mellitus or inborn errors of metabolism) where a strict metabolic control is an important part of the therapy, often result in not only insufficient supply with vitamins and minerals, but also lower amounts of saturated and polyunsaturated fatty acids are provided by the diet.

The omega-6 essential fatty acid, linoleic acid (LA; 18:2 n-6) is present in large amounts in vegetable seed oils such as sunflower seed oil, while the omega-3 essential fatty acid, alpha-linolenic acid (ALA; 18:3 n-3) is present in large concentrations in flaxseed oil and green, leafy vegetables. However, the long-chain metabolites are also present in their preformed state in the nature. The omega-6 arachidonic acid (AA; 20:4n-6) is present in lean red meat and chicken, in egg yolks, while the omega-3 PUFA, eicosapentaenoic acid (EPA or 20:5n-3) and docosahexaenoic acid (DHA; 22:6n-3) are synthesized in abundance by marine algae and therefore are present in concentrated form in cold water fish and in marine oils. In non-fish eaters, eggs are an important source of DHA and dairy products provide some EPA (Bradbury 2011).

The nutritional role of prebiotics

Prebiotics are selectively fermented dietary ingredients, which promote specific changes in the composition and/or activity of the gastrointestinal microbiota and thus confer benefits on the host health (Roberfroid et al. 2010). The prebiotic concept was first defined by Gibson and Roberfroid in 1995 and updated one decade later (Roberfroid et al. 2010, Gibson 2004). Accordingly, food ingredients are classified as prebiotic when they fulfill three main criteria: firstly, they escape digestion in the upper gastrointestinal tract and reach the colon intact; secondly, they are fermented by the intestinal microflora; and thirdly, they selectively stimulate the growth and/or activity of those intestinal bacteria which are associated with health and wellbeing (Moro et al. 2002, Haarman and Knol 2005, Schmelzle et al. 2003).

The role of prebiotics was investigated most extensively in the field of infant nutrition (Boehm et al 2003, Thomas et al 2010). Oligosaccharides were demonstrated to rep-

resent the most important dietary factor in human milk, promoting the development of beneficial intestinal flora (Bertino et al 2012, Barile 2013).

Prebiotics mimic the beneficial functional properties of human milk oligosaccharides: they increase stool colony counts of bifidobacteria and lactobacilli (Moro et al. 2002, Haarman and Knol 2005, Schmelzle et al. 2003); moreover, they inhibit the adhesion of pathogen organisms and interact with immune cells (Jeurink et al. 2013, Seifert and Watzl 2007). In bottle-fed infants prebiotic supplementation results in stools similar to those of breastfed neonates (i.e. leads to change in consistency to softer stools, a higher stool frequency and similar faecal pH) (Moro et al. 2002, Fanaro et al. 2005, Ashley et al 2012).

Because of their complex structure, oligosaccharides with a structure identical to that of human milk oligosaccharides (HMOs) are not yet available for supplementation purposes; however, oligosaccharides of other origin were tested for their bifidogenic effect in humans. At present galactooligosaccharides (GOS) and fructooligosaccharides (FOS) are considered the most relevant prebiotic oligosaccharides used in supplementation studies.

The main natural sources of prebiotics are chicory, Jerusalem artichoke, garlic, leek, onion and asparagus.

2. Methodical aspects of conducting a systematic review and meta-analysis

A systematic review is a review of a clearly formulated question that uses systematic and explicit methods to identify, select and critically appraise relevant research and to collect and analyze data from the studies that are included in the review (Moher et al. 2009). Its key characteristics according to the Cochrane Handbook are (Higgins and Green 2011):

- a clearly stated set of objectives with pre-defined eligibility criteria for studies
- an explicit, reproducible methodology
- a systematic search that attempts to identify all studies that would meet the eligibility criteria
- an assessment of the validity of the findings of the included studies (for example through the assessment of risk of bias)
- a systematic presentation, and synthesis, of the characteristics and findings of the included studies.

Systematic reviews and meta-analyses become increasingly important in the previous decades in health care: they are the primary source for clinicians to keep their knowledge up-to-date and they are also used as a starting point when developing clinical practice guidelines.

2.1 The PRISMA Statement

The value of systematic reviews depends on the way it was conducted, on the quality of collected studies and on the clarity of reporting. In 1996 an international group developed guideline (the QUOROM Statement = Quality of Reporting of Meta-Analyses) to optimize the reporting of meta-analyses; this guideline was revised and renamed in 2009: the PRISMA Statement (Preferred Reporting Items for Systematic reviews and Meta-Analyses) is practically a check-list which contains the items which

are important to be included when reporting a systematic review; the PRISMA statement aims to help both review authors and reviewers to improve the quality of reporting (Moher et al 2009; Liberati et al. 2009).

2.2 Framing the question and deciding on important outcomes

Framing the question covers following issues: a) the patient population, b) the intervention of interest, c) the comparator and d) the outcomes of interest has to be specified thoughtfully (Guyatt et al. GRADE 2. 2011). In systematic reviews investigating not the effect of an intervention and therefore summarizing not randomized controlled trials (RCTs), framing the question may not include all these issues. A very important decision by framing the question is to decide how widely the patients and the intervention should be defined: across the included patients and interventions the magnitude of effect on the key outcomes should be the same to avoid misleading estimates for some subpopulations or some subgroups of interventions (Guyatt et al. GRADE 2. 2011).

2.3. Searching for studies, selecting studies and collecting data

The Cochrane Central Register of Controlled Trials (CENTRAL), MEDLINE and EMBASE are the most often searched databases. Besides, trials registers are an increasingly important source of information. Both free-text and subject headings (for example Medical Subject Headings (MeSH)) should be used for search. The Cochrane Organization developed highly sensitive search strategies (filters) which facilitate the identification of randomized trials in the MEDLINE database (Lefebvre et al. 2011).

Eligibility of studies should be assessed and data extraction from study reports should be done by at least two independent reviewers. Studies, rather than reports, are the units of interest, therefore, one publication may contain more studies; on the other hand, the same study may be described and published in multiple reports (Higgins and Deeks 2011).

2.4 Assessment of the validity of the findings

A bias is a systematic error, which can lead to either underestimation or overestimation of the true intervention effect. Differences in risks of bias can help explaining the variation among results (the heterogeneity) of the studies included in a systematic review. The main types of "risk of bias" in case of a randomized controlled study are: selection bias (lack of allocation concealment/lack of randomization: those enrolling patients know about the group to which the next enrolled patient will be allocated), performance bias (the study personnel and/or included subjects are not blinded), detection bias (blinding of outcome assessment), attrition bias (incomplete outcome data addressed) and reporting bias (the reporting of some outcomes is incomplete or absent, while others are reported in detail in the Results section)(Higgins et al. 2011; Guyatt et al. GRADE 4. 2011).

2.5 Data synthesis and meta-analysis

Meta-analysis is the statistical combination of results from two or more separate studies. First, the type of data for the outcome measurements has to be identified. Five main types of data can be differentiated:

- dichotomous data, where outcomes are one of two possible categorical responses;
- 2. continuous data, where outcomes can take any value on a numerical scale;
- 3. ordinal data, where the outcome is one of several ordered categories;
- 4. *counts and rates* calculated from counting the number of events that each individual experiences;
- 5. *time-to-event* (typically survival) *data* that analyze the time until an event occurs, but where not all individuals in the study experience the event .

Effect measures for continuous outcomes

The **mean difference (MD),** or more correctly, the difference in means, measures the absolute difference between the mean values in two groups. The **standardized mean difference (SMD)** is used when the studies all assess the same outcome but measure it in a variety of ways, for example on different scales. In this case it is necessary to standardize the results of these studies to an uniform scale before they can be com-

$$SMD = \frac{\text{Difference in mean outcome between groups}}{\text{Standard deviation of outcome among participants}} \, .$$

Effect measures for counts and rates

The rate ratio (RR) compares the rate of events in the two groups by dividing one by the other. The rate difference, which measures the difference in rates, is used less common as a summary statistic (Deeks et al. 2011).

Meta-analysis is typically a two-stage process. In the first stage, the *effect measure* (summary statistic) is calculated for each study (for example the difference between means in case of continuous data, rate ratio in case of rates). In the second stage, a *summary (pooled) effect estimate* is calculated as a weighted average of the effects estimated in the individual studies. A weighted average is defined as weighted average = $\frac{\text{sum of (estimate } \times \text{weight)}}{\text{sum of weights}} = \frac{\sum Y_i W_i}{\sum W_i}$

where Y_i is the intervention effect estimated in the i study, W_i is the weight given to the i study, and the summation is across all studies. The bigger the weight given to the i study, the more it will contribute to the weighted average. For ratio measures (for example rate ratio), Yi is the natural logarithm of the measure.

Two types of statistical models can be used for combination of effect estimates across studies: a *random-effects meta-analysis* is preferred, in case the studies are not all estimating the same intervention effect (i.e. there is heterogeneity among studies that cannot readily be explained); while in the case each study is estimating

exactly the same quantity (i.e the effect in both magnitude and direction is the same value in every study) a *fixed-effect meta-analysis* is performed.

Heterogeneity

Heterogeneity means any kind of variability among studies in a systematic review, including clinical (variability in the participants), methodological (variability in study design and risk of bias) and statistical diversity (variability in the intervention effects). In meta-analyses we often use the I^2 -test for quantifying heterogeneity. First, a chi-squared (χ^2 , or Chi²) test is conducted to assess whether observed differences in results can be explained by chance alone. A low P value indicates heterogeneity of intervention effects. The I^2 test is a transformation of the Chi² test (Q):

$$I^2 = \left(\frac{Q - df}{Q}\right) \times 100\%$$

where df is the degree of freedom for the Chi² test (Higgins and Thompson 2002).

I² test describes the percentage of variability in point estimates that is due to heterogeneity. I² test 0% to 40% may be interpreted as a level of heterogeneity which is not important, 30% to 60% may represent moderate, 50% to 90% may represent substantial, while 75% to 100% may represent considerable heterogeneity. The advantage of the test is that it does not depend on the number of studies included into the metanalysis.

Subgroup analyses may be done as a means of investigating heterogeneous results (for example for different age groups of participants).

2.6 Presenting results

Forest plots are often used for visualizing results of the meta-analysis (**Figure 2**). A forest plot presents effect estimates and confidence intervals for both individual studies and meta-analyses (Lewis and Clarke 2001). The area of the block, representing point estimates, indicates the weight of that study in the meta-analysis while the horizontal line illustrates the confidence interval, usually with a 95% level of confi-

dence. The confidence interval indicates whether each intervention effect was individually statistically significant.

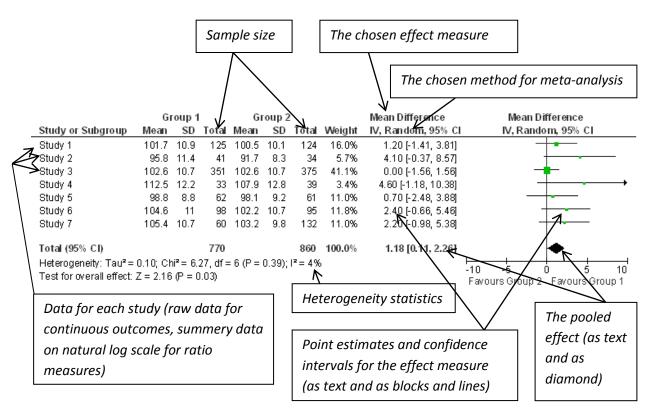


Figure 2. Details provided in a forest plot

3. Aims

- 1. Sex hormones may influence the activity of enzymes playing a role in the synthesis of long-chain polyunsaturated fatty acids. Our aim was to systematically review the gender-specific differences between women and men in the contribution of LCPUFAs to the fatty acid composition of the lipid pools most often reported in the literature for the characterization of fatty acid status, i.e. plasma phospholipids (PL), plasma cholesteryl esters (CE), plasma triacylglycerols (TG), total plasma lipids, erythrocyte and platelet lipids as well as subcutaneous adipose tissue from the abdomen or buttock.
- 2. The mainstream of phenylketonuria management is lifelong restriction of protein intake; however, this dietary restriction may be accompanied with insufficient dietary intake of long-chain polyunsaturated fatty acids. Our aim was to assess whether significant depletion of LCPUFA can be detected in phenylketonuria (PKU) patients on diet and whether LCPUFA supplementation is an effective way to increase the availability of LCPUFA in PKU patients.
- 3. Acute respiratory tract infections are the most common reason for people to seek medical help in developed countries. Strategies to prevent acute infections include the administration of different immunostimulants, vitamins and trace elements; recently, also probiotics were described to be effective in reducing the incidence rate of upper respiratory tract infections and that of diarrhea. In the case of probiotic supplementation, a large number (billions) of living probiotic bacteria have to be administered on a daily basis to ensure the continuous colonisation of the intestine and reach the desired health benefits; prebiotics may be an alternative and easier way to reach the same positive effects that usually are subscribed to probiotics. Our objective was to systematically review the available literature and to assess the efficacy of prebiotics in the prevention of acute infectious diseases in the pediatric age group.

4. Meta-analyses

4.1. Gender differences in long-chain polyunsaturated fatty acid status of healthy subjects: a systematic review and meta-analysis

4.1.1. Inclusion criteria

To be included into the review, a study needed to meet all of the following characteristics: 1. a study carried out in humans, 2. at least 14 participants included, 3. n–3 or n–6 LCPUFA status is reported in both males and females, 4. healthy individuals with normal weight were included, or population-based surveys was carried out in that the majority of participants were considered healthy, 5. omnivorous participants were included, 6. there was no dietary intervention (especially no lipid modified diet) or drug therapy before sample collection, 7. investigators measured at least 12 fatty acids by gas-liquid chromatography so the percentage distribution data contained the principal fatty acids of the fatty acid spectrum and presumably reflected realistic proportion of fatty acids in the given lipid fraction.

4.1.2. Search strategy

Electronic searches

Ovid MEDLINE (www.ovid.com), Scopus (www.scopus.elsevier.com), and the Cochrane Library CENTRAL database (www.thecochranelibrary.org) were searched from inception to February 2011 for studies containing LCPUFA values of both men and women (boys and girls) using text terms with appropriate truncation and relevant indexing terms. The search was in the form [n–3 LCPUFA terms] or [n–6 LCPUFA terms] and [biomarker terms] and [gender terms] and [differ*] and [human studies]. The results obtained by the full Ovid MEDLINE search strategy are shown in **Table1**. The searches of the two other databases were based also on this strategy. We did not apply any language restriction.

Table 1. Search strategy for Ovid MEDLINE from 1950 to February 2011

#	Searche History	Results
1	exp Fatty Acids/	317728
2	fatty acid.mp.	70673
3	fatty acids.mp.	130870
4	omega-6.mp.	2800
5	omega6.mp.	164
6	omega-3.mp.	8847
7	omega3.mp.	345
8	PUFA.mp.	4608
9	PUFAs.mp.	1865
10	LC-PUFA.mp.	183
11	LC-PUFAs.mp.	74
12	LCPUFA.mp.	257
13	LCPUFAs.mp.	101
14	polyunsaturated.mp.	18317
15	poly-unsaturated.mp.	253
16	1 or 2 or 3 or 4 or 5 or 6 or 7 or 8 or 9 or 10 or 11 or 12 or 13 or 14 or 15	359501
17	exp Docosahexaenoic Acids/	4233
18	"docosahexaenoic acid".mp.	5362
19	"docosahexanoic acid".mp.	147
20	"docosahexenoic acid".mp.	32
21	DHA.mp.	6023
22	exp Eicosapentaenoic Acid/	3411
23	"eicosapentaenoic acid".mp.	5329
24	"eicosapentanoic acid".mp.	120
25	"eicosapentenoic acid".mp.	10
26	EPA.mp.	6841
27	"arachidonic acid".mp.	33150
28	"linolenic acid".mp.	5411
29	"linoleic acid".mp.	11898
30	17 or 18 or 19 or 20 or 21 or 22 or 23 or 24 or 25 or 26 or 27 or 28 or 29	56012
31	16 or 30	371034
32	exp Plasma/	12829
33	plasma.mp.	603550
34	exp Serum/	57441
35	serum.mp.	698887
36	exp Erythrocytes/	142985
	erythrocyte.mp.	84213
38	erythrocytes.mp.	139866

39 "red blood cell".mp. 17930 40 "red blood cells".mp. 13721 41 RBC.mp. 5881 42 "RBCS".mp. 5881 43 "red blood corpuscles".mp. 17 44 "red blood corpuscles".mp. 111 45 exp Blood Platelets/ 60845 46 platelets.mp. 85591 47 platelets.mp. 85591 48 "thrombocytes".mp. 2736 49 "thrombocytes".mp. 3009 51 "granulocytes".mp. 41550 52 "granulocytes".mp. 27420 53 "peripheral blood mononuclear cell".mp. 26148 55 "PBMC".mp. 6225 57 "PBMC".mp. 6225 58 LDLmp. 625 59 exp Lipoproteins, LDL/ 3726 60 HDLmp. 48739 61 exp Adipocytes/ 10869 62 "adipocytes".mp. 9666 63 "adipocytes".mp. 9666 64 "exp Adipocytes, ADL/ 48739 65 "adipocytes, ADL/ 9666 65 "adipocytes".mp. 9966 66 "adipocytes".mp. 9966 <th></th> <th></th> <th></th>			
41 RBC.mp. 13721 42 "RBCs".mp. 5881 43 "red blood corpuscle".mp. 17 44 "red blood corpuscles".mp. 17 45 exp Blood Platelets/ 60845 46 platelet.mp. 153311 47 platelets.mp. 8559 48 "Hrombocyte".mp. 2736 49 "thrombocytes".mp. 3009 51 "granulocytes".mp. 41550 52 "granulocytes".mp. 2722 54 "peripheral blood mononuclear cell".mp. 202 55 "PBMCs".mp. 26148 56 "PBMCs".mp. 2625 57 exp Lipoproteins, LDL/ 37206 58 LDL.mp. 6225 59 exp Lipoproteins, LDL/ 37206 61 HDL.mp. 4873 62 "adipocytes".mp. 4873 63 "adipocytes".mp. 9666 64 "adipocytes".mp. 9666 65 "adipocytes".mp. 9676 66 "adipocytes".mp. 9686 67 "adipocytes".mp. 9686 68 "adipocytes".mp. 9686 69 "adipocytes".mp. 9686			17930
42 "RBCS".mp. 5881 43 "red blood corpuscles".mp. 17 44 "red blood corpuscles".mp. 111 45 exp Blood Platelets/ 60845 6 platelet.mp. 153311 47 platelets.mp. 85559 48 "thrombocyte".mp. 2736 49 "thrombocytes".mp. 3009 50 exp Granulocytes".mp. 107967 51 "granulocyte".mp. 41550 52 "granulocytes".mp. 27420 53 "peripheral blood mononuclear cells".mp. 26148 55 "PBMC".mp. 26148 56 "PBMC".mp. 2625 57 exp Lipoproteins, LDL/ 37206 8 LDL.mp. 4275 58 LDL.mp. 48739 60 HDL.mp. 48739 61 exp Adipocytes, HDL/ 30623 62 "adipocytes".mp. 9666 63 "adipocytes".mp. 9666 64 "exp Adipocytes".mp. 9666 65 "adipocytes".mp. 9666 66 "adipocytes".mp. 9666 67 "adipocytes".mp. 96866 68 "adipocytes".mp. 9686<			
43 "red blood corpuscle".mp. 17 44 "red blood corpuscles".mp. 111 45 ex plood Platelets/ 60845 46 platelet.mp. 153311 47 platelets.mp. 85559 48 "thrombocyte".mp. 3009 49 "thrombocytes".mp. 3009 50 exp Granulocytes/ 107967 51 "granulocytes".mp. 41550 52 "granulocytes".mp. 27420 54 "beripheral blood mononuclear cell".mp. 26148 55 "PBMC".mp. 26148 56 "PBMCS".mp. 6225 57 exp Lipoproteins, LDL/ 37206 58 LDL.mp. 61778 60 HDL.mp. 48739 61 exp Adipocytes/ 10869 62 "adipocytes".mp. 9666 63 "adipocytes".mp. 9666 64 exp Adipocytes ".mp. 19481 65 "adipocytes".mp. 6964 66 * adipocytes".mp. 6966 67 "adipocytes".mp. 6966 68 (male and female).mp. 69064 69 "adipocytes".mp. 69064 69 (males and females).mp.<		·	
44 "red blood corpuscles".mp. 111 45 exp Blood Platelets/ 60845 46 platelet.mp. 153311 47 platelets.mp. 85559 48 "thrombocyte".mp. 2736 49 "thrombocytes".mp. 3009 50 exp Granulocytes/ 107967 51 "granulocyte".mp. 41550 52 "granulocytes".mp. 27420 53 "peripheral blood mononuclear cell".mp. 2202 54 "peripheral blood mononuclear cells".mp. 26148 55 "PBMC".mp. 3326 56 "PBMCS".mp. 61778 57 exp Lipoproteins, LDL/ 37206 58 LDL.mp. 61778 59 exp Lipoproteins, HDL/ 30523 60 HDL.mp. 48739 61 exp Adipocytes/ 10869 62 "adipocytes".mp. 666 63 "adipocytes".mp. 6906 64 "ad jocytes".mp. 6906 65 "Bodipocytes".mp. 6906 66 "adjoose tissue".mp. 6906 67 "ad jocytes".mp. 6906 68 "adjoose tissue".mp. 6906 69 (male			
45 exp Blood Platelets/mp. 153311 47 platelet.mp. 153311 47 platelets.mp. 85559 48 "thrombocyte".mp. 2736 49 "thrombocytes".mp. 107967 51 "granulocytes".mp. 41550 52 "granulocytes".mp. 27420 53 "peripheral blood mononuclear cell".mp. 2202 54 "peripheral blood mononuclear cells".mp. 6148 55 "PBMC".mp. 13027 66 "PBMCs".mp. 61278 57 exp Lipoproteins, LDL/ 37206 58 LDL.mp. 48739 61 reb Adipocytes/ 10869 62 "adipocytes".mp. 9666 63 "adipocytes".mp. 60132 64 exp Adipocytes/ 10869 65 "adipocytes".mp. 60162 66 "adipocytes".mp. 60162 67 "adipocytes".mp. 60162 68 exp Adipocytes/ 60132			
46 platelet.mp. 153311 47 platelets.mp. 85559 48 "thrombocytes".mp. 3009 50 exp Granulocytes/ 107967 51 "granulocytes".mp. 27420 52 "granulocytes".mp. 27420 53 "peripheral blood mononuclear cell".mp. 2022 54 "peripheral blood mononuclear cells".mp. 26148 55 "PBMC".mp. 3087 66 "PBMCS".mp. 6225 57 exp Lipoproteins, LDL/ 37206 58 LDL.mp. 61778 59 exp Lipoproteins, HDL/ 30523 60 HDL.mp. 48739 61 exp Adipocytes/ 10869 62 "adipocytes".mp. 9666 63 "adipocytes".mp. 9666 64 "adipocytes".mp. 69064 65 "adipocytes".mp. 69064 66 "adipocytes".mp. 69064 67 "ali and 66 60132 68 (male and female).mp. 1802378 67			111
47 platelets.mp. 85559 48 "thrombocyte".mp. 2736 49 "thrombocytes".mp. 3009 50 exp Granulocytes/ 107967 51 "granulocytes".mp. 41556 52 "granulocytes".mp. 27420 53 "peripheral blood mononuclear cells".mp. 26148 55 "PBMC".mp. 13087 56 "PBMCs".mp. 6225 57 exp Lipoproteins, LDL/ 37206 58 LDL.mp. 61778 59 exp Lipoproteins, HDL/ 3053 60 HDL.mp. 48739 61 exp Adipocytes/ 10869 62 "adipocytes".mp. 9666 63 "adipocytes".mp. 9666 64 "adipocytes".mp. 9666 65 "adipocytes".mp. 9666 66 "adipocytes".mp. 9666 67 "adipocytes".mp. 9666 68 "adipocytes".mp. 9666 69 "adipocytes".mp. 96064 67 "alian de			60845
48 "thrombocyte".mp. 2736 49 "thrombocytes".mp. 3009 50 exp Granulocytes/ 107967 51 "granulocyte".mp. 41550 52 "granulocytes".mp. 27420 53 "peripheral blood mononuclear cell".mp. 2202 54 "peripheral blood mononuclear cells".mp. 13087 55 "PBMC".mp. 13087 6 "PBMCs".mp. 6225 7 exp Lipoproteins, LDL/ 37206 8 LDL.mp. 61778 9 exp Lipoproteins, HDL/ 30523 40 HDL.mp. 48739 61 exp Adipocytes/ 10869 62 "adipocytes".mp. 9666 63 "adipocytes".mp. 9666 64 exp Adipocytes".mp. 9666 65 "adipocytes".mp. 9666 66 "adipocytes".mp. 9666 67 "adipocytes".mp. 9604 68 "adipocytes".mp. 9004 69 "adipocytes".mp. 9004 61 exp Adipocytes".mp. 9004 62 "adipocytes".mp. 90064 7 31 and 66 105882 8 (male and females).mp.	46	platelet.mp.	
49 "thrombocytes".mp. 3009 50 exp Granulocytes/ 107967 51 "granulocyte".mp. 41550 52 "granulocytes".mp. 27420 53 "peripheral blood mononuclear cell".mp. 2202 54 "peripheral blood mononuclear cells".mp. 26148 55 "PBMC".mp. 13087 56 "PBMCs".mp. 6225 57 exp Lipoproteins, LDL/ 37206 58 LDL.mp. 61778 9 exp Lipoproteins, HDL/ 30523 60 HDL.mp. 48739 61 exp Adipocytes/ 10869 62 "adipocytes".mp. 9666 63 "adipocytes".mp. 9666 64 exp Adipose Tissue/ 60132 65 "adipose tissue".mp. 6904 66 "adipose tissue".mp. 69064 67 "3 I and 66 105882 68 (male and female).mp. 3607668 69 (males and females).mp. 120913 70 (men and women).mp. 153945 71 (boys and girls).mp. 31298 72 gender.mp. 313154 73 sex.mp. 478441 74 6 8 or 69 or 70 or 71 or 72 or 73 376383 7	47	platelets.mp.	85559
50 exp Granulocytes/ 107967 51 "granulocyte".mp. 41550 52 "granulocytes".mp. 27420 53 "peripheral blood mononuclear cell".mp. 26148 55 "PBMC".mp. 13087 56 "PBMCs".mp. 6225 57 exp Lipoproteins, LDL/ 37206 58 LDL.mp. 61778 59 exp Lipoproteins, HDL/ 30523 60 HDL.mp. 48739 61 exp Adipocytes/ 10869 62 "adipocytes".mp. 9666 63 "adipocytes".mp. 9666 63 "adipocytes".mp. 60132 65 "adipose tissue".mp. 60132 65 "adipose tissue".mp. 60164 66 "adipose tissue".mp. 60064 67 31 and 66 10582 68 (male and female).mp. 120913 69 (male and female).mp. 120913 71 (boys and girls).mp. 120913 72 gender.mp. 131454 73	48	"thrombocyte".mp.	2736
51 "granulocyte".mp. 41550 52 "granulocytes".mp. 27420 53 "peripheral blood mononuclear cell".mp. 2202 54 "peripheral blood mononuclear cells".mp. 26148 55 "PBMC".mp. 13087 66 "PBMCs".mp. 6225 75 exp Lipoproteins, LDL/ 37206 8 LDL.mp. 61778 59 exp Lipoproteins, HDL/ 30523 60 HDL.mp. 48739 61 exp Adipocytes/ 10869 62 "adipocytes".mp. 9666 63 "adipocytes".mp. 60132 64 exp Adipose Tissue/ 60132 65 "adipose tissue".mp. 60132 66 "adipose tissue".mp. 60132 67 "31 and 66 105882 68 (male and female).mp. 3607688 69 (males and females).mp. 120913 70 (men and women).mp. 153945 71 (boys and girls).mp. 131454 72 gender.mp. 478441 73 sex.mp. 478441 74 6 differ\$.mp. 3509153 75 and 76 75 and 76 75 and 76	49	"thrombocytes".mp.	3009
52 "granulocytes".mp. 27420 53 "peripheral blood mononuclear cell".mp. 2202 54 "peripheral blood mononuclear cells".mp. 26148 55 "PBMCs".mp. 6225 56 "PBMCs".mp. 6225 57 exp Lipoproteins, LDL/ 37206 58 LDL.mp. 61778 59 exp Lipoproteins, HDL/ 30523 60 HDL.mp. 48739 61 exp Adipocytes/ 10869 62 "adipocytes".mp. 9666 63 "adipocytes".mp. 60132 64 exp Adipose Tissue/ 60132 65 "adipose tissue".mp. 60132 66 "adipose tissue".mp. 60132 67 "al and 66 105882 68 (male and female).mp. 120913 69 (males and females).mp. 120913 70 (men and women).mp. 153945 71 (boys and girls).mp. 131454 72 gender.mp. <t< td=""><td>50</td><td>exp Granulocytes/</td><td>107967</td></t<>	50	exp Granulocytes/	107967
53 "peripheral blood mononuclear cell".mp. 2202 54 "peripheral blood mononuclear cells".mp. 26148 55 "PBMC".mp. 13087 56 "PBMCs".mp. 6225 57 exp Lipoproteins, LDL/ 37206 58 LDL.mp. 61778 59 exp Lipoproteins, HDL/ 30523 60 HDL.mp. 48739 61 exp Adipocytes/ 10869 62 "adipocytes".mp. 9666 63 "adipocytes".mp. 69064 64 exp Adipose Tissue/ 60132 65 "adipose tissue".mp. 69064 66 32 or 33 or 34 or 35 or 36 or 37 or 38 or 39 or 40 or 41 or 42 or 43 or 44 or 45 or 46 or 47 or 48 or 48 or 49 or 50 or 51 or 52 or 53 or 54 or 55 or 56 or 57 or 58 or 59 or 60 or 61 or 62 or 63 or 64 or 65 1802378 67 31 and 66 105882 68 (male and female).mp. 3607668 69 (males and females).mp. 120913 70 (men and women).mp. 153945 71 (boys and girls).mp. 31298 72 gender.mp. 3736338 </td <td>51</td> <td>"granulocyte".mp.</td> <td>41550</td>	51	"granulocyte".mp.	41550
54 "peripheral blood mononuclear cells".mp. 26148 55 "PBMC".mp. 13087 56 "PBMCs".mp. 6225 57 exp Lipoproteins, LDL/ 37206 58 LDL.mp. 61778 59 exp Lipoproteins, HDL/ 30523 60 HDL.mp. 48739 61 exp Adipocytes/ 10869 62 "adipocyte".mp. 9666 63 "adipocytes".mp. 69064 64 exp Adipose Tissue/ 60132 65 "adipose tissue".mp. 69064 66 "3 1 and 66 105882 67 31 and 66 105882 68 (male and female).mp. 3607668 69 (males and females).mp. 120913 70 (men and women).mp. 153945 71 (boys and girls).mp. 31298 72 gender.mp. 313154 73 sex.mp. 478441 74 68 or 69 or 70 or 71 or 72 or 73 3736338 75 67 and 74 23001 76 differ\$.mp. 3509153 77 5 and 76 75 and 76	52	"granulocytes".mp.	27420
55 "PBMC".mp. 13087 56 "PBMCs".mp. 6225 57 exp Lipoproteins, LDL/ 37206 58 LDL.mp. 61778 59 exp Lipoproteins, HDL/ 30523 60 HDL.mp. 48739 61 exp Adipocytes/ 10869 62 "adipocyte".mp. 9666 63 "adipocytes".mp. 60132 64 exp Adipose Tissue/ 60132 65 "adipose tissue".mp. 60964 66 "adipose tissue".mp. 69064 67 31 and 66 105882 68 (male and female).mp. 3607668 69 (males and females).mp. 120913 70 (men and women).mp. 153945 71 (boys and girls).mp. 31298 72 gender.mp. 313454 73 sex.mp. 478441 74 68 or 69 or 70 or 71 or 72 or 73 373638 75 75 and 74 23001 76 differ\$.mp. 3509153 75 75 and 76	53	"peripheral blood mononuclear cell".mp.	2202
56 "PBMCS".mp. 6225 57 exp Lipoproteins, LDL/ 37206 58 LDL.mp. 61778 59 exp Lipoproteins, HDL/ 30523 60 HDL.mp. 48739 61 exp Adipocytes/ 10869 62 "adipocyte".mp. 9666 63 "adipocytes".mp. 60132 65 "adipose tissue".mp. 60132 65 "adipose tissue".mp. 69064 66 32 a cor 33 or 34 or 35 or 36 or 37 or 38 or 39 or 40 or 41 or 42 or 43 or 44 or 45 or 46 or 47 or 48 or 49 or 50 or 51 or 52 or 53 or 54 or 55 or 56 or 57 or 58 or 59 or 60 or 61 or 62 or 63 or 64 or 65 1802378 67 31 and 66 105882 68 (male and female).mp. 3607668 69 (males and females).mp. 120913 70 (men and women).mp. 153945 71 (boys and girls).mp. 31298 72 gender.mp. 313454 73 sex.mp. 478441 74 68 or 69 or 70 or 71 or 72 or 73 3736338 75 67 and 74 23001 76 differ\$.mp. 3509153 77 5 and 76 753 and 76	54	"peripheral blood mononuclear cells".mp.	26148
57 exp Lipoproteins, LDL/ 37206 58 LDL.mp. 61778 59 exp Lipoproteins, HDL/ 30523 60 HDL.mp. 48739 61 exp Adipocytes/ 10869 62 "adipocytes".mp. 9666 63 "adipocytes".mp. 60132 65 "adipose tissue".mp. 69064 65 "adipose tissue".mp. 69064 66 32 or 33 or 34 or 35 or 36 or 37 or 38 or 39 or 40 or 41 or 42 or 43 or 44 or 45 or 46 or 47 or 48 or 49 or 50 or 51 or 52 or 53 or 54 or 55 or 56 or 57 or 58 or 59 or 60 or 61 or 62 or 63 or 64 or 65 1802378 67 31 and 66 105882 68 (male and female).mp. 3607668 69 (males and females).mp. 120913 70 (men and women).mp. 153945 71 (boys and girls).mp. 31298 72 gender.mp. 478441 73 sex.mp. 478441 74 68 or 69 or 70 or 71 or 72 or 73 3736338 75 67 and 74 23001 76 differ\$.mp. 3509153 77 <td< td=""><td>55</td><td>"PBMC".mp.</td><td>13087</td></td<>	55	"PBMC".mp.	13087
58 LDL.mp. 61778 59 exp Lipoproteins, HDL/ 30523 60 HDL.mp. 48739 61 exp Adipocytes/ 10869 62 "adipocyte".mp. 9666 63 "adipocytes".mp. 60132 65 "adipose tissue".mp. 69064 66 32 or 33 or 34 or 35 or 36 or 37 or 38 or 39 or 40 or 41 or 42 or 43 or 44 or 45 or 46 or 47 or 48 or 49 or 50 or 51 or 52 or 53 or 54 or 55 or 56 or 57 or 58 or 59 or 60 or 61 or 62 or 63 or 64 or 65 1802378 67 31 and 66 105882 68 (male and female).mp. 3607668 69 (males and females).mp. 120913 70 (men and women).mp. 153945 71 (boys and girls).mp. 31298 72 gender.mp. 3736338 73 sex.mp. 478441 74 68 or 69 or 70 or 71 or 72 or 73 3736338 75 67 and 74 23001 76 differ\$.mp. 3509153 77 75 and 76 7543	56	"PBMCs".mp.	6225
59 exp Lipoproteins, HDL/ 30523 60 HDL.mp. 48739 61 exp Adipocytes/ 10869 62 "adipocyte".mp. 9666 63 "adipocytes".mp. 19481 64 exp Adipose Tissue/ 60132 65 "adipose tissue".mp. 69064 66 32 or 33 or 34 or 35 or 36 or 37 or 38 or 39 or 40 or 41 or 42 or 43 or 44 or 45 or 46 or 47 or 48 or 49 or 50 or 51 or 52 or 53 or 54 or 55 or 56 or 57 or 58 or 59 or 60 or 61 or 62 or 63 or 64 or 65 1802378 67 31 and 66 105882 68 (male and female).mp. 3607668 69 (males and females).mp. 120913 70 (men and women).mp. 153945 71 (boys and girls).mp. 31298 72 gender.mp. 31454 73 sex.mp. 478441 74 68 or 69 or 70 or 71 or 72 or 73 3736338 75 67 and 74 23001 76 differ\$.mp. 3509153 77 75 and 76 7543	57	exp Lipoproteins, LDL/	37206
60 HDL.mp. 48739 61 exp Adipocytes/ 10869 62 "adipocyte".mp. 9666 63 "adipocytes".mp. 19481 64 exp Adipose Tissue/ 60132 65 "adipose tissue".mp. 69064 66 32 or 33 or 34 or 35 or 36 or 37 or 38 or 39 or 40 or 41 or 42 or 43 or 44 or 45 or 46 or 47 or 48 or 49 or 50 or 51 or 52 or 53 or 54 or 55 or 56 or 57 or 58 or 59 or 60 or 61 or 62 or 63 or 64 or 65 1802378 67 31 and 66 105882 68 (male and female).mp. 3607668 69 (males and females).mp. 120913 70 (men and women).mp. 153945 71 (boys and girls).mp. 31298 72 gender.mp. 31454 73 sex.mp. 478441 74 68 or 69 or 70 or 71 or 72 or 73 3736338 75 67 and 74 23001 76 differ\$.mp. 3509153 77 75 and 76 7543	58	LDL.mp.	61778
61 exp Adipocytes/ 62 "adipocyte".mp. 9666 63 "adipocytes".mp. 19481 64 exp Adipose Tissue/ 65 "adipose tissue".mp. 69064 66 32 or 33 or 34 or 35 or 36 or 37 or 38 or 39 or 40 or 41 or 42 or 43 or 44 or 45 or 46 or 47 or 48 or 49 or 50 or 51 or 52 or 53 or 54 or 55 or 56 or 57 or 58 or 59 or 60 or 61 or 62 or 63 or 64 or 65 67 31 and 66 68 (male and female).mp. 3607668 69 (males and females).mp. 120913 70 (men and women).mp. 153945 71 (boys and girls).mp. 31298 72 gender.mp. 31298 73 sex.mp. 478441 74 68 or 69 or 70 or 71 or 72 or 73 75 and 76 75 and 76	59	exp Lipoproteins, HDL/	30523
62 "adipocyte".mp. 9666 63 "adipocytes".mp. 19481 64 exp Adipose Tissue/ 60132 65 "adipose tissue".mp. 69064 66 32 or 33 or 34 or 35 or 36 or 37 or 38 or 39 or 40 or 41 or 42 or 43 or 44 or 45 or 46 or 47 or 48 or 49 or 50 or 51 or 52 or 53 or 54 or 55 or 56 or 57 or 58 or 59 or 60 or 61 or 62 or 63 or 64 or 65 67 31 and 66 105882 68 (male and female).mp. 3607668 69 (males and females).mp. 120913 70 (men and women).mp. 153945 71 (boys and girls).mp. 31298 72 gender.mp. 131454 73 sex.mp. 478441 74 68 or 69 or 70 or 71 or 72 or 73 3736338 75 67 and 74 23001 76 differ\$.mp. 3509153 77 75 and 76	60	HDL.mp.	48739
63 "adipocytes".mp.1948164 exp Adipose Tissue/6013265 "adipose tissue".mp.6906466 32 or 33 or 34 or 35 or 36 or 37 or 38 or 39 or 40 or 41 or 42 or 43 or 44 or 45 or 46 or 47 or 48 or 49 or 50 or 51 or 52 or 53 or 54 or 55 or 56 or 57 or 58 or 59 or 60 or 61 or 62 or 63 or 64 or 65180237867 31 and 6610588268 (male and female).mp.360766869 (males and females).mp.12091370 (men and women).mp.15394571 (boys and girls).mp.3129872 gender.mp.13145473 sex.mp.47844174 68 or 69 or 70 or 71 or 72 or 73373633875 67 and 742300176 differ\$.mp.350915377 75 and 767543	61	exp Adipocytes/	10869
64 exp Adipose Tissue/ 65 "adipose tissue".mp. 66 32 or 33 or 34 or 35 or 36 or 37 or 38 or 39 or 40 or 41 or 42 or 43 or 44 or 45 or 46 or 47 or 48 or 49 or 50 or 51 or 52 or 53 or 54 or 55 or 56 or 57 or 58 or 59 or 60 or 61 or 62 or 63 or 64 or 65 67 31 and 66 68 (male and female).mp. 69 (males and females).mp. 70 (men and women).mp. 71 (boys and girls).mp. 72 gender.mp. 73 sex.mp. 74 68 or 69 or 70 or 71 or 72 or 73 75 and 76 75 and 76 75 and 76 75 31 and 76 75 32 or 33 or 34 or 35 or 36 or 37 or 38 or 39 or 40 or 41 or 42 or 43 or 44 or 45 or 46 or 47 or 48 or 48 or 48 or 49 or 50 or 51 or 52 or 53 or 54 or 55 or 56 or 57 or 58 or 59 or 60 or 61 or 62 or 63 or 64 or 65 1802378 180	62	"adipocyte".mp.	9666
65 "adipose tissue".mp. 66 32 or 33 or 34 or 35 or 36 or 37 or 38 or 39 or 40 or 41 or 42 or 43 or 44 or 45 or 46 or 47 or 48 or 49 or 50 or 51 or 52 or 53 or 54 or 55 or 56 or 57 or 58 or 59 or 60 or 61 or 62 or 63 or 64 or 65 67 31 and 66 68 (male and female).mp. 69064 69 (males and females).mp. 70 (men and women).mp. 120913 71 (boys and girls).mp. 72 gender.mp. 131454 73 sex.mp. 1478441 74 68 or 69 or 70 or 71 or 72 or 73 75 and 76 75 and 76 75 and 76	63	"adipocytes".mp.	19481
6632 or 33 or 34 or 35 or 36 or 37 or 38 or 39 or 40 or 41 or 42 or 43 or 44 or 45 or 46 or 47 or 48 or 49 or 50 or 51 or 52 or 53 or 54 or 55 or 56 or 57 or 58 or 59 or 60 or 61 or 62 or 63 or 64 or 6518023786731 and 6610588268(male and female).mp.360766869(males and females).mp.12091370(men and women).mp.15394571(boys and girls).mp.3129872gender.mp.13145473sex.mp.4784417468 or 69 or 70 or 71 or 72 or 7337363387567 and 742300176differ\$.mp.35091537775 and 767543	64	exp Adipose Tissue/	60132
49 or 50 or 51 or 52 or 53 or 54 or 55 or 56 or 57 or 58 or 59 or 60 or 61 or 62 or 63 or 64 or 65 67 31 and 66 (male and female).mp. 69 (males and females).mp. 120913 70 (men and women).mp. 153945 71 (boys and girls).mp. 31298 72 gender.mp. 313454 73 sex.mp. 478441 74 68 or 69 or 70 or 71 or 72 or 73 75 and 76 3509153 77 75 and 76	65	"adipose tissue".mp.	69064
68 (male and female).mp.360766869 (males and females).mp.12091370 (men and women).mp.15394571 (boys and girls).mp.3129872 gender.mp.13145473 sex.mp.47844174 68 or 69 or 70 or 71 or 72 or 73373633875 67 and 742300176 differ\$.mp.350915377 75 and 767543	66		1802378
69 (males and females).mp. 70 (men and women).mp. 153945 71 (boys and girls).mp. 72 gender.mp. 73 sex.mp. 74 68 or 69 or 70 or 71 or 72 or 73 75 and 76 120913 153945 131454 131454 131454 13786338 13786338 13786338 13786338 13786338 13786338 13786338 13786338 13786338	67	31 and 66	105882
70 (men and women).mp. 153945 71 (boys and girls).mp. 31298 72 gender.mp. 131454 73 sex.mp. 478441 74 68 or 69 or 70 or 71 or 72 or 73 3736338 75 67 and 74 23001 76 differ\$.mp. 3509153 77 75 and 76 7543	68	(male and female).mp.	3607668
71 (boys and girls).mp. 31298 72 gender.mp. 131454 73 sex.mp. 478441 74 68 or 69 or 70 or 71 or 72 or 73 3736338 75 67 and 74 23001 76 differ\$.mp. 3509153 77 75 and 76 7543	69	(males and females).mp.	120913
72 gender.mp. 131454 73 sex.mp. 478441 74 68 or 69 or 70 or 71 or 72 or 73 3736338 75 67 and 74 23001 76 differ\$.mp. 3509153 77 75 and 76 7543	70	(men and women).mp.	153945
73 sex.mp. 478441 74 68 or 69 or 70 or 71 or 72 or 73 3736338 75 67 and 74 23001 76 differ\$.mp. 3509153 77 75 and 76 7543	71	(boys and girls).mp.	31298
74 68 or 69 or 70 or 71 or 72 or 73 3736338 75 67 and 74 23001 76 differ\$.mp. 3509153 77 75 and 76 7543	72	gender.mp.	131454
75 67 and 74 76 differ\$.mp. 77 75 and 76 23001 3509153 77 75 and 76	73	sex.mp.	478441
76 differ\$.mp. 3509153 77 75 and 76 7543	74	68 or 69 or 70 or 71 or 72 or 73	3736338
77 75 and 76 7543	75	67 and 74	23001
	76	differ\$.mp.	3509153
78 limit 77 to humans 6309	77	75 and 76	7543
	78	limit 77 to humans	6309

4.1.3. Data extraction

Titles and abstracts found by the electronic and bibliographic searches were screened for inclusion by a single reviewer (SL). Because there were a large number of papers which were present in more than one databases, duplications were filtered out to compile the final list of titles and abstracts to be screened. Thereafter it was checked which of the titles and abstracts meet the predetermined eligibility criteria. If articles seemed to meet the inclusion criteria, or the title and the abstract left room for doubt, the full text of the article was evaluated by two independent reviewers (SL and KF). If the two reviewers disagreed about the eligibility, the study was discussed in detail to reach a consensus decision.

Data for each study included were extracted by a single reviewer (SL) into a Microsoft Office Excel 2007 database file. To provide a standardized format, units of measurement were recalculated to percentage contribution of LCPUFA to total fatty acid composition of the relevant tissue (% weight/weight) from original data in the publication. If it was not possible to convert data, we tried to contact the authors. If the authors couldn't be contacted (in most of the cases because of the long time elapsed since the publication of the papers), or the data were not available in the original form any more, those studies were excluded. The original forms in which fatty acid data in the included studies were expressed are shown in Table 1. In some studies the investigators measured the fatty acid composition in plasma, in other studies in serum. Because we feel that there is no major difference in the percentage fatty acid composition of plasma and serum, for the uniformity of discussion, we use the term 'plasma' throughout the text.

4.1.4. Statistical analysis

Statistical analyses were performed using the Review Manager 5.1 Software (The Cochrane Collaboration, Oxford, United Kingdom). Mean differences (MD) was used for the analysis of continuous data. The confidence interval (CI) was established at 95%. P values of less than 0.05 were considered to indicate statistical significance.

Statistical heterogeneity was assessed using the I^2 statistics (I^2 of 50% or more indicating presence of heterogeneity).

4.1.5. Results

Study inclusion

The flow diagram of the literature search for this review is shown on Figure 3.

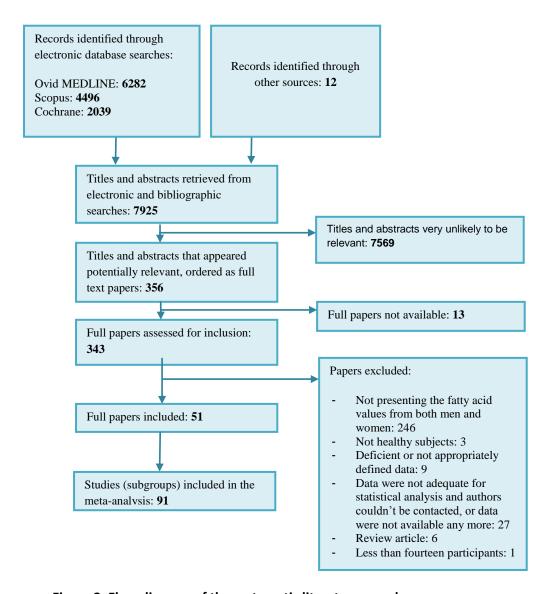


Figure 3. Flow diagram of the systematic literature search

Methods used in the studies reviewed

Among the studies reviewed there were some methodological differences in the analytical methods used for the determination of fatty acid composition of the different biomarkers. The blood samples collected were stored in a deep frozen state, but the storage temperature was different (-20°C, -30 °C, -40 °C, -70 °C or -80 °C). Lipid extraction was carried out by chloroform and methanol in most, but not all the studies. The separation of different lipid fractions was performed by thin layer chromatography. Fatty acid analysis was carried out by gas-liquid chromatography in all studies. In some studies a packed column was used instead of capillary columns (capillary column: 34 papers, packed column: 10 papers, column type not reported: 7 papers)

Biomarkers identified

We found 11 publications analyzing plasma PLs, whereas 8 analyzed plasma CEs, 5 plasma TGs, 18 total plasma lipids, 9 total erythrocyte membrane lipids, 1 platelets and 7 adipose tissue fatty acid composition. A description of biomarkers identified in three or more studies, including also the number of studies, subgroups, participants from both gender, and the results of the primary analysis (MD, I²) is presented in **Table 3**. We discuss in detail only these biomarkers. Descriptive data of biomarkers detected in less than three studies are presented in **Table 2**.

Fatty acids reported

In this study we focused on the following 8 PUFAs: LA, gamma-linolenic acid (GLA), dihomo-gamma-linolenic acid (DHGLA) and AA from the n-6 series and ALA, eicosapentaenoic acid (EPA), docosapentaenoic acid (DPA) and DHA from the n-3 series.

Table 2: Basic characteristics of included studies

			Participant cha	racteristics		Biomarkers reported ¹		
First author, publication year	Country	No. of included male subjects	No. of included female subjects	Age of males	Age of females		Fatty acids reported	Original expression of data
Ando, 1990 [5]	Japan	31 ^a 29 37	68 45 22	54.3±7.4 y 59.8±4.9 y 64.9±7.3 y	57.0±8.6 y 59.0±6.0 y 68.3±8.8 y	plasma total lipids	LA, DHGLA, AA, EPA, DHA	mg/dl (mean, SD)
Antonini, 1970 [6]	Italy	11	11	29±6.6 y	27.4±6.2 y	adipose tissue	LA	w/w% (mean, SD)
Araki, 1990 [7]	Japan	27 ^b 57	110 121	20–49 y 50–79 y	20–49 y 50–79 y	plasma total lipids	LA, AA, EPA, DHA	w/w% (mean, SD)
Bakewell, 2006 [8]	United King- dom	13	23	26±5 y	23±4 y	plasma total lipids, plasma TG, plasma FFA, plasma PC, plasma CE	LA, GLA, DHGLA, AA, ALA, EPA, DPA, DHA	w/w % (mean, SD)
Bolton-Smith, 1997 [9]	Scotland	2308	2049	40–59 y	40–59 y	adipose tissue	LA, GLA, DHGLA, AA	w/w% (mean, SD)
Brouwer, 1997 [10]	The Nether- lands (Cura- cao)	51	26	56±8 y	58±5 y	plasma CE	LA, GLA, DHGLA, AA, ALA, EPA, DHA	mol% (mean, SD)
Cheng, 2003 [11]	Taiwan	10 ^c 10 ^d	10 10	10–11 y 10–11 y	10-11 y 10-11 y	plasma TG	LA	w/w% (mean, SEM)
Christensen, 1999 [12]	Denmark	35	25	38±10 y	38±11 y	granulocytes, platelet	LA, AA, ALA, EPA, DPA, DHA	w/w% (mean, SD)
di Giuseppe, 2009 [13]	Italy, Belgium, United King- dom	50	224	47±1.1 y	44±0.5 y	plasma total lipids, erythrocyte membrane	ALA, EPA, DPA, DHA	w/w% (geometric mean, SEM or 95% CI)
Elizondo-Montemayor, 2010 [14]	Mexico	49	51	6–12 y	6–12 y	plasma PL	LA, GLA, DHGLA, AA, ALA, EPA, DPA, DHA	w/w% (mean, SD)
Fernandez-Real, 2001 [15]	Spain	38	40	40.1±13.3 y	38.1±9.3 y	plasma total lipids	LA, DHGLA, AA, EPA, DHA	w/w% (mean, SD)
Geppert, 2010 [16]	United King- dom	40	34	32.6±8.0 y	32.7±7.3 y	platelet PC, platelet PEA	LA, GLA, DHGLA, AA, ALA, EPA, DPA, DHA	w/w% (mean, SD)
Giltay, 2004 [17]	The Nether- lands	72	71	29.6±12.9 y	27.4±10.2 y	plasma CE	LA, GLA, DHGLA, AA, ALA, EPA, DHA	w/w% (mean, 95% CI)
Glew, 2010 [18]	Nigeria	22	29	55.5±13.5 y	47.6±8.3 y	plasma PL	LA, GLA, DHGLA, AA, ALA, EPA, DPA, DHA	w/w% (mean, SD)

Glew, 2002 [19]	Nigeria	37	36	14 y	13 y	plasma PL	LA, GLA, DHGLA, AA, ALA, EPA, DPA, DHA	w/w% (mean, SD)
Hagenfeldt, 1975 [20]	Sweden	6	8	26–35 y	23–31 y	plasma FFA	LA	w/w% (mean, SEM)
Hirai, 2005 [21]	Japan	76	76	68.6±10.6 y	67.8±11.2 y	plasma total lipids	LA, GLA, DHGLA, AA, ALA, EPA, DPA, DHA	μg/ml (mean, SD)
Hirai, 2000 [22]	Japan, The Netherlands	33 ^f 20 ^g	29 19	university stu- dents	university stu- dents	plasma total lipids	LA, AA, ALA, EPA, DHA	mg/100ml (mean, SD)
Hodge, 2007 [23]	Australia, United King- dom, Italy, Greece	2048	2391	55±8.8 y	54.4±8.5 y	plasma PL	LA, AA, ALA, EPA, DHA	w/w% (mean, SD)
Innis, 1988 [24]	Canada	11 ^h 9 ^h 41 ^h 14 ^h 12 ⁱ	17 12 59 13 12	11–15 y 16–20 y 21–50 >50 y 21–50 y	11–82 y 16–20 y 21–50 >50 y 21–50 y	erythrocyte PC, erythrocyte PEA	LA, GLA, DHGLA, AA, ALA, EPA, DPA, DHA	w/w% (mean, SEM)
Iwamoto, 2002 [25]	Japan	20	20	adults	adults	plasma CE	LA, AA, EPA, DHA	mol% (mean, SEM)
Jagannathan, 1969 [26]	India	27	15	22–50 y	26–45 y	adipose tissue	LA	w/w% (mean, SEM)
Kale, 2008 [27]	India	25	21	33.6±9.7 y	35.1±8.0 y	erythrocyte membrane	LA, GLA, AA, ALA, EPA, DPA, DHA	w/w% (mean, SD)
Karlsson, 2006 [28]	Sweden	6	9	11.3–15.4 y	11.7–14.5 y	plasma PL	LA, GLA, DHGLA, AA, ALA, EPA, DHA	mol% (mean, SD)
Kieu, 2002 [29]	South Vi- etnam	32 ^d 40 ^e 39 ^c	68 58 59	47.5±5.5 y 46.6±5.0 y 46.4±4.7 y	47.5±5.2 y 47.6±6.4 y 47.2±5.9 y	plasma total lipids	LA, GLA, DHGLA, AA, ALA, EPA, DPA, DHA	w/w% (mean, SD)
Kuriki, 2003 [30]	Japan	15	79	45.3±10.6 y	47.2±8.1 y	plasma total lipids	LA, AA, ALA, EPA, DPA, DHA	w/w% (mean, SD)
Lee, 2000 [31]	Hong Kong	81	113	43.4±11.2 y	43.8±11.6 y	plasma total lipids	LA, GLA, AA, ALA, EPA, DHA	w/w% (mean, SD)
Lemaitre, 2008 [32]	Israel	112 ^b 49 44	118 42 52	<45 y 45–60 y ≥60 y	<45 y 45–60 y ≥60 y	erythrocyte membrane	LA, DHGLA, AA, EPA, DHA	w/w% (mean, SD)
Lucas, 2009 [33]	Canada	127	170	18-74 y	18–74 y	plasma PL	LA, AA, ALA, EPA, DPA, DHA	w/w% (mean, SD)
Mamalakis, 2006 [34]	Greece	59	71	37.7±7.9 y	36.2±6.7 y	adipose tissue	LA, GLA, DHGLA, AA, ALA, EPA, DPA, DHA	w/w% (mean, SD)

Mamalakis, 1998 [35]	Greece	85	59	23–69 y	23–69 у	adipose tissue	LA, DHGLA, AA, ALA, EPA, DPA, DHA	w/w% (mean, SD)
McNamara, 2010 [36]	USA	10	10	35.9±8.8 y	36.1±9.3 y	erythrocyte membrane	LA, DHGLA, AA, EPA, DPA, DHA	w/w% (mean, SEM)
Melchert, 1987 [37]	Germany	38	70	21–77 y	18-85 y	plasma total lipids, HDL	LA, GLA, DHGLA, AA, ALA, DHA	w/w% (mean, SD)
						plasma CE	LA, GLA, DHGLA, AA, ALA, EPA, DHA	
						plasma TG	LA, GLA, AA, ALA, DHA	
						plasma FFA	LA, DHGLA, AA, ALA, DHA	
Metherel, 2009 [38]	Canada	9	7	22.4±1.2 y	22.1±1.8 y	whole blood, plasma total lipids, erythrocyte mem- brane, fingertip prick blood	LA, GLA, DHGLA, AA, ALA, EPA, DPA, DHA	w/w% (mean, SD)
Mitchell, 1983 [39]	New Zealand	9	9	10–13 y	10–13 y	erythrocyte membrane	LA, DHGLA, AA, ALA, EPA, DPA, DHA	w/w% (mean, SD)
Nakamura, 1995 [40]	Japan	18 ^b	13	30–39 y	30–69 y	plasma total lipids	LA, AA, EPA, DHA	w/w% (mean, SD)
		13	13	40–49 y	40–49 y			
		12	15	50-59 y	50-59 y			
		12	14	60–69 y	60–69 y			
Nikkari, 1995 [41] Finlar	Finland	41	41	43±4.3 y	40±4.3 y	plasma PL	LA, GLA, DHGLA, AA, EPA, DHA	w/w% (mean, SD)
						plasma CE	LA, GLA, DHGLA, AA, ALA, EPA, DHA	
						plasma TG	LA, AA, ALA, DHA	

Saadatian-Elahi, 2009	Greece	91 ^j	100	45–64 y	45–64 y	plasma PL	LA, GLA, DHGLA, AA,	w/w% (mean, SD)
[42]	Spain	93	100				ALA, EPA, DPA, DHA	
	Spain	96	100					
	Spain	94	100					
	Italy	90	99					
	Italy	94	99					
	Italy	96	100					
	Germany	95	96					
	Germany	96	99					
	Netherlands	95	100					
	UK	95	100					
	Denmark	96	100					
	Sweden	100	95					
	Sweden	94	99					
Sfar, 2010 [43]	Tunesia	96	104	55.7±13.1 y	53.1±13.7 y	plasma total lipids	LA, AA, ALA, EPA, DHA	w/w% (mean, SD)
Smit, 2003 [44]	The Nether- lands	29	34	22–49 y	22–47 y	erythrocyte membrane	LA, GLA, DHGLA, AA, ALA, EPA, DPA, DHA	mol% (mean, SD)
Sutherland, 1995 [45]	Fiji	39 ^c	44	39±16 y	36±15 y	erythrocyte membrane	LA, GLA, DHGLA, AA,	w/w% (mean, SD)
	•	37 ^d	34	38±14 y	43±18 y		ALA, EPA, DPA, DHA	
Takita, 1996 [46]	Japan	28 ^b	15	20–29 y	20–29 y	plasma total lipids	LA, AA, EPA, DHA	w/w% (mean, SD)
		87	58	30-39 y	30–39 y			
		81	48	40–49 y	40–49 y			
		20	30	50-59 y	50-59 y			
		17	10	60–69 y	60–69 y			
Tavendale, 1992 [47]	Scotland	529 ^b	518	40–44 y	40–44 y	adipose tissue	LA, GLA, DHGLA, AA,	w/w% (mean, SD)
		508	469	45–49 y	45–49 y		DPA, DHA	
		593	479	50-54 y	50-54 y			
		555	463	55-59 y	55–59 y			
Tjønneland, 1993 [48]	Denmark	23	63	42–63 y	40–63 y	adipose tissue	LA, GLA, DHGLA, AA, ALA, EPA, DHA	w/w% (mean, SD)
Umemura, 2005 [49]	Japan	175	246	19.4±1 y	19.2±0.5 y	plasma total lipids	LA, GLA, DHGLA, AA, ALA, EPA, DPA, DHA	w/w% (mean, SD
Vallés, 1988 [50]	Spain	49	49	16–75 у	16–75 y	plasma PL, plasma TG, plasma FFA, plasma CE, platelet PL, platelet TG, platelet FFA	LA, EPA	w/w% (mean, SD)
Warensjö, 2006 [51]	Sweden	554	295	40.6±9.1 y	40.6±9.9 y	plasma CE	LA, GLA, DHGLA, AA, ALA, EPA, DHA	w/w% (mean, SD)
Wennberg, 2011 [52]	Sweden	308	248	53±7.7 y	58±7.2 y	plasma PL	ALA, EPA, DPA, DHA	w/w% (mean, rang

Wennberg, 2007 [53]	Sweden	288	207	55.2±7.5 y	55.4±7.6 y	plasma PL, erythrocyte membrane	ALA, EPA, DPA, DHA	w/w% (median, SD)
Yamada, 2000 [54]	Japan	107 ^k 78 ^l	154 124	30–89 y	30–89 y	plasma total lipids	LA, AA, ALA, EPA, DPA, DHA	w/w% (mean, SEM)
Yeh, 1996 [55]	Nigeria	110 ^m 126 ⁿ	65 96	42.4±8.9 y 39.7±10.3 y	41.4±8.1 y 35.4±9.1 y	plasma total lipids	LA, GLA, DHGLA, AA, ALA, EPA, DHA	mol% (mean, SD)

Abbreviations:

^{*,} mean±SD; **, range.

a, groups classified by cluster analysis; b, groups classified by age range; c, urban region; d, rural region; e, suburban region; f, Japanese group; g, Dutch group; h, Inuit group classified by age range; i, Vancouver group; j, groups classified by geographic areas (Athens, Granada, Murcia, North Spain, Ragusa/Naples, Florence, Varese/Turin, Heidelberg, Potsdam, The Netherlands, Cambridge, Denmark, Malmö, Umeå); k, fishing group; l, farming group; m, senior staff group; n, junior staff group.

LA, linoleic acid; GLA, gamma-linolenic acid; DHGLA, dihomo-gamma-linolenic acid; AA, arachidonic acid; ALA, alpha-linolenic acid; EPA, eicosapentaenoic acid; DPA, docosapentaenoic acid, TG, triacylglycerol; PC, phosphatidylcholine; CE cholesteryl ester; PEA, phospatidylethanolamine; FFA, free fatty acids; PL, phospholipid.

Table 3: Primary analysis of biomarkers identified in three or more studies

		No. of indi-	No. of p	articipants	MD [95% CI]	Heterogenity
		vidual studies	Men	Women		(I ²)
total plasma lipids	LA	32	1594	2012	-1.55 [-1.96, -1.14] ^a	40%
пріцэ	GLA	11	738	881	-0.33 [-0.07, 0.00]	78%
	DHGLA	14	792	943	-0.05 [-0.17, 0.06]	92%
	AA	32	1594	2012	-0.22 [-0.37, -0.07] ^a	60%
	ALA	18	1137	1614	-0.03 [-0.06, 0.00]	60%
	EPA	32	1607	2166	0.02 [-0.07, 0.11]	69%
	DPA	11	634	1118	0.06 [0.03, 0.09] ^b	55%
	DHA	33	1644	2236	-0.12 [-0.22, -0.03] ^a	46%
plasma phospho- lipids	LA	22	3704	4163	-0.19 [-0.43, 0.05]	50%
	GLA	19	1480	1553	0.00 [-0.00, 0.01]	36%
	DHGLA	19	1480	1553	-0.07 [-0.15, 0.01]	56%
	AA	21	3655	4114	-0.42 [-0.65, -0.18] ^a	81%
	ALA	22	4052	4403	-0.00 [-0.01, 0.00]	72%
	EPA	24	4146	4493	-0.03 [-0.08, 0.02]	54%
	DPA	20	2002	2003	0.01 [-0.01, 0.04]	77%
	DHA	23	4097	4444	-0.37 [-0.51, -0.24] ^a	79%
olasma choles-	LA	8	805	530	-1.03 [-1.84, -0.21] ^a	45%
teryl esters	GLA	6	736	461	0.08 [0.03, 0.14] ^b	32%
	DHGLA	6	736	461	0.06 [0.02, 0.09] ^b	49%
	AA	7	756	481	0.21 [-0.24, 0.66]	79%
	ALA	6	736	461	-0.01 [-0.03, 0.00]	0%
	EPA	8	805	530	0.01 [-0.07, 0.10]	55%
	DHA	7	756	481	0.02 [-0.08, 0.04]	68%
plasma triacyl-	LA	6	128	138	0.35 [-0.81, 1.53]	57%
glycerols	AA	3	59	69	0.07 [-0.13, 0.28]	76%
	ALA	3	59	69	-0.03 [-0.20, 0.13]	0%
	DHA	3	59	69	-0.12 [-0.57, 0.32]	68%
erythrocyte	LA	10	363	371	0.05 [-0.19, 0.30]	0%
membrane lipids	GLA	5	139	140	-0.00 [-0.01, 0.01]	8%

	DHGLA	9	338	350	-0.02 [-0.12, 0.08]	18%
	AA	10	363	371	-0.41 [-0.86, 0.05]	64%
	ALA	8	480	577	0.01 [-0.02, 0.04]	69%
	EPA	12	701	802	0.03 [-0.00, 0.07]	27%
	DPA	9	495	590	0.14 [-0.05, 0.33]	79%
	DHA	12	698	802	-0.19 [-0.31, -0.06] ^a	3%
erythrocyte	LA	5	87	113	-0.11 [-2.61, 2.38]	39%
phosphatidyl- choline	GLA	5	87	113	0.09 [-0.23, 0.41]	43%
	DHGLA	5	87	113	-0.18 [-0.40, 0.04]	0%
	AA	4	46	54	-1.88 [-4.05, 0.29]	0%
	ALA	3	61	88	-0.01 [-0.42, 0.40]	0%
	EPA	5	87	113	-0.41 [-0.74, -0.08] ^a	0%
	DPA	5	87	113	0.32 [-0.05, 0.69]	0%
	DHA	5	87	113	-0.25 [-1.30, 0.80]	62%
erythrocyte phosphatidyl-	LA	5	87	113	0.26 [-0.58, 1.09]	0%
ethanolamine	GLA	4	78	101	0.02 [-0.13, 0.18]	0%
	DHGLA	5	87	113	-0.12 [-0.37, 0.13]	25%
	AA	5	87	113	0.12 [-2.08, 1.85]	38%
	ALA	5	87	113	0.08 [-0.15, 0.32]	0%
	EPA	5	87	113	0.01 [-1.10, 1.13]	30%
	DPA	5	87	113	0.25 [-0.38, 0.88]	6%
	DHA	5	87	113	0.21 [-1.16, 1.57]	0%
adipose tissue	LA	10	4698	4197	-0.49 [-0.60, -0.38] ^a	0%
	GLA	7	4575	4112	0.03 [0.01, 0.04] ^b	92%
	DHGLA	8	4660	4171	-0.04 [-0.05, -0.03] ^a	93%
	AA	8	4660	4171	0.03 [0.02, 0.05] ^b	74%
	ALA	3	167	193	0.01 [-0.02, 0.03]	0%
	EPA	3	167	193	0.01 [0.01, 0.02] ^b	0%
	DPA	6	2329	2059	-0.01 [-0.04, 0.02]	97%
	DHA	7	2352	2122	-0.01 [-0.04, 0.01]	96%

Abbreviations: a, significantly higher (p < 0.05) in women than in men; b, significantly higher (p < 0.05) in men than in women.

Fatty acid composition of total plasma lipids

Primary analysis showed significantly higher contribution of the n-6 essential fatty acid, LA and the n-6 long-chain metabolite, AA to plasma total lipids of women compared to men (Figure 4, Table 3).

As to n-3 fatty acids, the values of the principal LCPUFA, DHA were significantly higher (**Figure 5**), while the values of its precursor, DPA were significantly lower in women compared to men (**Table 3**). However, with the exception of LA and DHA, considerable heterogeneity was seen among the results of the individual studies.

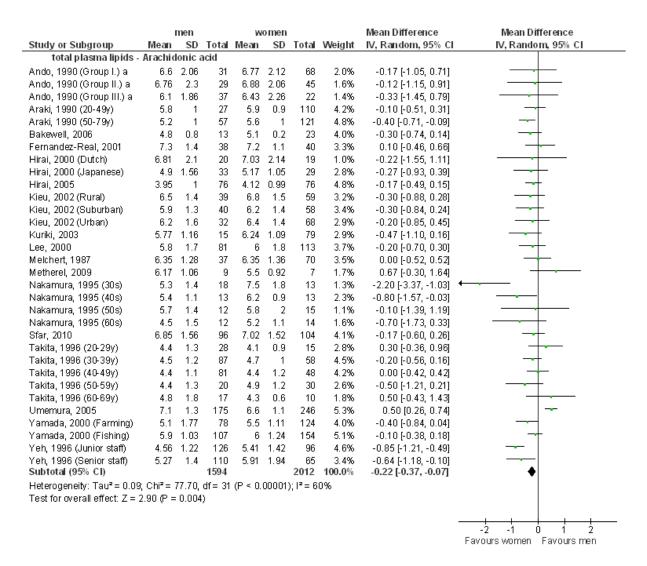


Figure 4. Mean difference in the percent contribution of AA to total plasma lipids of healthy male and female subjects

		men		W	omen			Mean Difference	Mean Difference
Study or Subgroup	Mean		Total		SD	Total	Weight	IV, Random, 95% CI	IV, Random, 95% CI
total plasma lipids -	Docosa	hexae	noic a	cid					
Ando, 1990 (Group I.) a	4.76	2.06	31	4.99	1.86	68	1.1%	-0.23 [-1.08, 0.62]	
Ando, 1990 (Group II.) a	4.46	2.16	29	4.54	1.76	45	0.9%	-0.08 [-1.02, 0.86]	
Ando, 1990 (Group III.) a	4.52	2.4	37	5.17	2.16	22	0.6%	-0.65 [-1.84, 0.54]	
Araki, 1990 (20-49y)	5.2	1.2	27	5.1	1.1	110	2.5%	0.10 [-0.40, 0.60]	
Araki, 1990 (50-79y)	5.8	1	57	5.6	0.9	121	4.5%	0.20 [-0.11, 0.51]	+-
Bakewell, 2006	1.2	0.3	13	1.7	0.7	23	4.2%	-0.50 [-0.83, -0.17]	
diGiuseppe, 2009	1.89	0.47	50	2.08	0.51	224	7.2%	-0.19 [-0.34, -0.04]	-
Fernandez-Real, 2001	2.03	0.4	38	1.95	0.7	40	5.3%	0.08 [-0.17, 0.33]	+
Hirai, 2000 (Dutch)	1.26	0.81	20	1.36	0.68	19	2.7%	-0.10 [-0.57, 0.37]	
Hirai, 2000 (Japanese)	2.2	1.22	33	2.22	0.95	29	2.2%	-0.02 [-0.56, 0.52]	
Hirai, 2005	3,69	1.43	76	3.38	1	76	3.4%	0.31 [-0.08, 0.70]	
Kieu, 2002 (Rural)	3	0.5	39	3.2	0.6	59	5.9%	-0.20 [-0.42, 0.02]	-
Kieu, 2002 (Suburban)	2.5	0.5	40	2.7	0.6	58	5.9%	-0.20 [-0.42, 0.02]	-
Kieu, 2002 (Urban)	2.2	0.4	32	2.5	0.5	68	6.5%	-0.30 [-0.48, -0.12]	-
Kuriki, 2003	4.05	0.99	15	4.6	0.83	79	2.3%	-0.55 [-1.08, -0.02]	
Lee, 2000	3.2	2.1	81	3.4	2.2	113	1.8%	-0.20 [-0.81, 0.41]	 -
Melchert, 1987	2.15	0.53	37	2.36	0.59	70	5.9%	-0.21 [-0.43, 0.01]	-
Metherel, 2009	1.07	0.26	9	1.34	0.24	7	5.4%	-0.27 [-0.52, -0.02]	-
Nakamura, 1995 (30s)	7.3	2.7	18	7.1	2.3	13	0.3%	0.20 [-1.57, 1.97]	
Nakamura, 1995 (40s)	7.2	3.2	13	7.4	2	13	0.2%	-0.20 [-2.25, 1.85]	
Nakamura, 1995 (50s)	8	2.8	12	7.2	1.3	15	0.3%	0.80 [-0.92, 2.52]	
Nakamura, 1995 (60s)	7.7	2	12	7.9	2.6	14	0.3%	-0.20 [-1.97, 1.57]	
Sfar, 2010	2.88	0.95	96	3.31	0.81	104	5.4%	-0.43 [-0.68, -0.18]	-
Takita, 1996 (20-29y)	2.5	1	28	2.4	0.6	15	2.6%	0.10 [-0.38, 0.58]	
Takita, 1996 (30-39y)	3.3	1.3	87	3.2	1.3	58	3.0%	0.10 [-0.33, 0.53]	
Takita, 1996 (40-49y)	3.6	1.3	81	3.9	1.2	48	2.9%	-0.30 [-0.74, 0.14]	
Takita, 1996 (50-59y)	4	1.6	20	4.1	0.9	30	1.3%	-0.10 [-0.87, 0.67]	
Takita, 1996 (60-69y)	4.3	1.6	17	3.7	1.2	10	0.7%	0.60 [-0.46, 1.66]	
Umemura, 2005	3.4	0.9	175	3.2	1	246	6.5%	0.20 [0.02, 0.38]	 -
Yamada, 2000 (Farming)	8.2	3.53	78	7.4	2.23	124	1.0%	0.80 [-0.08, 1.68]	
Yamada, 2000 (Fishing)	8.6	3.1	107	8.3	2.49	154	1.5%	0.30 [-0.41, 1.01]	
Yeh, 1996 (Junior staff)	4.11	1.5	126	4.52	1.44	96	3.5%	-0.41 [-0.80, -0.02]	
Yeh, 1996 (Senior staff) Subtotal (95% CI)	4.2	1.5	110 1644	4.51	1.83	65 2236	2.3% 100.0%	-0.31 [-0.84, 0.22] - 0.12 [-0.22, -0.03]	•
Heterogeneity: Tau2 = 0.03;	Chi² = 5	59.61.	df= 32	(P = 0.1)	002); ř	² = 46%	5		
Test for overall effect: $Z = 2$,,,				
									-2 -1 0 1 2

Figure 5. Mean difference in the percent contribution of DHA to total plasma lipids of healthy male and female subjects

To filter out the potential effect of age, we tried to classify the individual studies into three age categories: a) 0 to12 years, b) 13 to 50 years, and c) 51 years or more. In the case of plasma total lipids classification according to age categories yielded sufficient number of studies to evaluate the effect of gender in the two adult groups only. Among the long-chain metabolites, in the 13 to 50 years group only the values of DHA (MD: -0.16; 95% CI: -0.26, -0.06; 2418 participants; $I^2 = 45\%$) were significantly higher in women, while the values of DPA were significantly lower in women compared to men (MD: 0.07; 95% CI: 0.03, 0.01; 1130 participants; $I^2 = 67\%$). In the > 51 years age

group (containing postmenopausal women) the values of the n-6 LCPUFAs, DHGLA (MD: -0.14; 95% CI: -0.24, -0.04; 137 participants; $I^2 = 19$ %) and AA (MD: -0.25; 95% CI: -0.43, -0.08; 892 participants; $I^2 = 0$ %) were significantly higher in women compared to men, whereas DHA values did not differ between the two genders.

To consider the potential effect of diet, we classified the studies based on the results of Meyer BJ (2011) according to fish eating habits, assigning the Inuit of Nunavik and the Japanese as high n-3 LCPUFA intake group and all other people as low n-3 LCPUFA intake group. In the high fish (n-3 LCPUFA) consuming group values of AA were significantly higher in women compared to men (MD: -0.23; 95% CI: -0.44, -0.02; 2291 participants; $I^2 = 68\%$), while there was no significant difference in the DHA levels between the two groups (MD: -0.00; 95% CI: -0.12, 0.12; 2291 participants; $I^2 = 17\%$). In the low fish consuming group both AA (MD: -0.24; 95% CI: -0.41, -0.07; 1383 participants; $I^2 = 8\%$) and DHA (MD: -0.24; 95% CI: -0.31, -0.16; 1598 participants; $I^2 = 11\%$) were significantly higher in women.

Fatty acid composition of plasma phospholipids

Plasma PL compositional data were reported for the largest number of men (n = 4097) and women (n = 4444). There was one cross-sectional study among the included articles (Saadatian-Elahi 2009), that contained 16 subgroups stratified according to geographic areas; we were able to include 14 subgroups (in one area only women were recruited, whereas another subgroup had to be excluded, because there were also vegans among the participants).

Primary analysis revealed significantly higher contribution of both AA (MD: -0.42; 95% CI: -0.65, -0.18; 7769 participants; $I^2 = 81\%$) and DHA (MD: -0.37; 95% CI: -0.51, -0.24; 8541 participants; $I^2 = 79\%$) to the fatty acid composition of plasma PLs in women (**Figures 6 and 7**), while in LA and ALA there was no gender difference. There were not enough studies to carry out subgroup analysis either by age or fish eating habits.

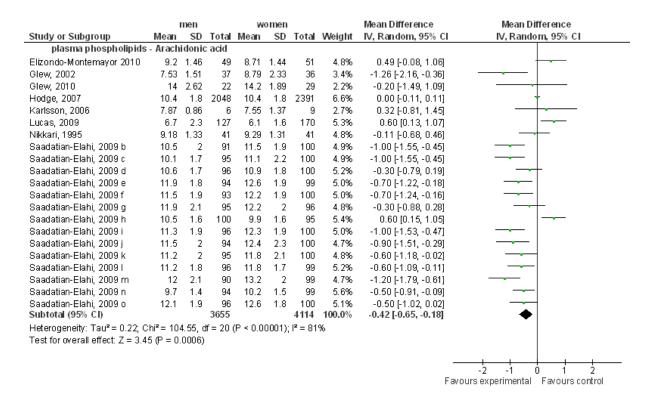


Figure 6. Mean difference in the percent contribution of AA to plasma PL of healthy male and female subjects

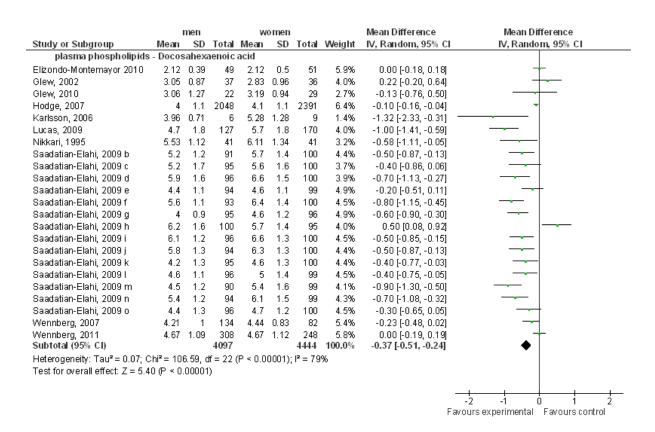


Figure 7. Mean difference in the percent contribution of DHA to plasma PL of healthy male and female subjects

Fatty acid composition of plasma cholesteryl esters

Plasma CE fatty acids were reported in 8 publications. There was no gender difference in AA and DHA values (**Table 3**), but GLA and DHGLA were found significantly higher in men as compared to women.

Fatty acid composition of plasma triacylglycerols

Five publications reported fatty acid composition of plasma TG in both men and women. The primary analysis showed no difference between the two sexes in any of the fatty acids discussed (**Table 3**).

Fatty acid composition of erythrocyte membrane total lipids

There were 9 publications reporting fatty acid composition of the erythrocyte membrane total lipids in both sexes. The primary analysis showed significant difference only in DHA values, which were higher in women than in men (Table 2). Because most of these studies were carried out in participants older than 50 years, in spite of the apparent abundance of data (12 studies with 1224 participants), there was no possibility to carry out subgroup analysis either by age or fish eating habits.

Fatty acid composition of erythrocyte phosphatidylcholine and phosphatidylethanolamine

There was one paper reporting erythrocyte phosphatidylcholine and phosphatidylethanolamine fatty acid composition in 5 subgroups [24]. In erythrocyte PC the values of EPA were significantly higher in women than in men, whereas in erythrocyte PEA there was no difference between the two sexes in any of the fatty acids discussed (**Table 3**).

Fatty acid composition of adipose tissue

In adipose tissue the values of LA and DHGLA were significantly higher in women than in men, while the values of GLA, AA, and from the n-3 series the values from EPA were significantly higher in men than in women.

4.1.6. Discussion

The endogenous synthesis of AA and DHA from their essential fatty acid precursors, LA and ALA require the contribution of elongases and desaturases. Since the elongation steps are rapid, whereas the desaturation steps are slower, the latter are considered as the rate limiting steps. In this systematic review we found higher contributions of the LCPUFAs AA and DHA to both plasma total lipids and plasma PL in women than in men. We also found significantly higher values of DHA, but not that of AA in women than in men in erythrocyte lipids. It may be assumed with good reason that the higher AA and DHA values found in plasma PLs of women may be due to the higher activity of desaturases, especially that of delta-6-desaturase.

The effects of sex hormones on essential fatty acid metabolism in humans have already been reviewed in detail by Childs et al. (Childs et al. 2008). In short, human studies demonstrated that males and females differ in their ability to synthesize n-3 LCPUFA from ALA, leading to the higher circulating concentrations of DHA in women than in men (Childs et al. 2008). In the same review significant relationship between plasma and tissue fatty acid composition and circulating sex hormone concentrations was seen, suggesting that estrogen stimulates, whereas testosterone inhibits the conversion of essential fatty acids into their longer-chain metabolites. This is consistent with findings of both animal studies (Extier et al. 2010) and human stable-isotope studies, which also showed that women have a higher capacity than men to synthesize DHA from ALA (Burdge and Wootton 2002; Pawlosky et al. 2003).

We also tried to evaluate the influence of age on the gender differences in fatty acid composition of biological samples. Since previously several authors suggested that gender differences seen in fatty acid compositional data may be due to the higher estrogen levels in women, we decided to make subgroups according to the presumably changing estrogen levels of women in the course of their life cycle (a) 0-12 years, b) 13-50 years and c) 50-75 years). In serum total lipids the higher levels of LCPUFAs in women than in men were also seen in the oldest age group, where mostly postmenopausal women were present. This finding on its own may indicate that not only the higher estrogen levels are responsible for the differences observed between sexes in LCPUFA values; e.g. beside the physiological and hormonal changes caused by ageing, different age groups can have very different dietary habits, which may also influence the composition of serum lipids. Moreover, many postmenopausal women receive estrogen supplementation therapy, which fact may further modify the picture. The data obtained in the present study indicate that gender differences in fatty acid status may be relevant also in the elderly.

Diet is considered as the major factor influencing fatty acid composition of tissues. Plasma PL fatty acid composition represents the dietary intake of fatty acids over periods of weeks or months before sample collection (Arab and Akbar 2002; Ma et al. 1995), while the rate of changes in red blood cells is slower than that seen in plasma lipids Katan et al. 1991; Glatz et al. 1989). The adipose tissue fatty acid pattern represents the diet ingested in the previous one to two and half years (Plakké et al. 1983; Beynen et al. 1980). In the vast majority of the studies included into our review the composition of diets was not investigated, so it is not possible to tell to what extent dietary fatty acids influence our results. However, there were enough studies to carry out a subgroup analysis in the total plasma lipid fraction by n-3 LCPUFA intake characteristic for the investigated populations. This analysis resulted in an appreciable decrease of the degree of heterogeneity, which confirms the important role of diet in determining LCPUFA status. In the high DHA consuming group (including Japanese and Inuit subjects) no significant difference between men and women was seen in DHA values. This observation suggests that gender is a significant potential confounding variable mainly in populations with low dietary n-3 intakes.

We think that our systematic review has some strongpoints. Firstly, we were able to identify a considerable number of studies investigating a relatively great number of women and men. Secondly, the studies included into the present review originated from a wide diversity of geographic location; consequently, the results obtained may be applied without serious geographical restriction. Thirdly, the studies analyzed were carried out during more than three decades, so the phenomena observed do not seem to be changing over time. However, there are also some weaknesses of our study. Firstly, studies included were mostly observational studies and not randomized controlled trials, so the quality control potential of the studies included into the review was limited. Secondly, in case of plasma PL 14 data sets (subgroups) originated from the same cross-sectional study, so these data influenced the statistical results notably. However, these subgroups represented 14 different geographic areas, and so there are good arguments for handling them as different studies. Thirdly, the analytical methods used by the different research groups were not standardized. However, methods of fatty acid analysis are not yet as rigorously standardized as many other laboratory methods, so any systematic review on fatty acid data faces the same difficulty. It is also important to add that there are several factors, which influence fatty acid metabolism and differ between genders (e.g. dietary fatty acid intake, alcohol ingestion, relative body fatness or level of physical activity). These factors may all contribute to the gender differences observed in fatty acid composition of different biological samples; however, the main objective of the present review was not to investigate the potential contribution of these factors, but to draw attention to the fact that gender-related differences exist.

In supplementation studies reporting fatty acid composition in serum PL, serum total lipids or erythrocyte membrane lipids, gender distribution should be regarded as significant potential confounding variable.

4.2. Long-chain polyunsaturated fatty acid status in patients with phenylketonuria as compared to healthy controls: a systematic review and meta-analysis

4.2.1. Inclusion criteria

To be included into the review, a study was required to meet the following characteristics: 1. be a study on patients of any age with PKU on low-protein diet and 2. be a case-control study reporting n–3 and/or n–6 LCPUFA status in patients and in healthy controls; or randomized controlled trial (RCT) investigating the effect of LCPUFA supplementation on LCPUFA status of patients with PKU.

4.2.2. Search strategy

(www.ovid.com), Scopus (www.scopus.elsevier.com), MEDLINE (www.bireme.br) the Cochrane Library **CENTRAL** database and (www.thecochranelibrary.org) were searched from inception to February 2011. The search was repeated in a reduced form in May 2012. Text terms with appropriate truncation and relevant indexing terms were used to identify articles. The search was in the form [n-3 LCPUFA terms] or [n-6 LCPUFA terms] and [PKU terms]. The Ovid MEDLINE search strategy is shown in **Table 4**. The search of the three other databases was based also on this strategy. The electronic search was supplemented with articles in the reference lists of the relevant studies and review articles. We did not apply any language restriction.

Table 4. Search strategy for Ovid MEDLINE from inception to May 2012

#	Search History	Results
1	exp Fatty Acids/	334085
2	fatty acid\$.mp.	171724
3	omega-6.mp.	3218
4	omega6.mp.	231
5	omega-3.mp.	10617
6	omega3.mp.	447
7	PUFA\$.mp.	6794
8	LC-PUFA\$.mp.	286
9	LCPUFA\$.mp.	383
10	polyunsaturated.mp.	20783
11	poly-unsaturated.mp.	306
12	exp Docosahexaenoic Acids/	4792
13	docosahexaenoic acid.mp.	6268
14	DHA.mp.	7110
15	exp Eicosapentaenoic Acid/	3740
16	eicosapentaenoic acid.mp.	6069
17	EPA.mp.	8452
18	arachidonic acid.mp.	34948
19	linoleic acid.mp.	13132
20	linolenic acid.mp.	6117
21	1 or 2 or 3 or 4 or 5 or 6 or 7 or 8 or 9 or 10 or 11 or 12 or 13 or 14 or $$	399834
	15 or 16 or 17 or 18 or 19 or 20	
22	exp Phenylketonurias/	5958
23	PKU.mp.	1984
24	phenylketonuria\$.mp.	6700
25	hyperphenylalaninaemia.mp.	254
26	HPA.mp.	9639
27	22 or 23 or 24 or 25 or 26	16339
28	21 and 27	313

4.2.3. Data extraction

Two reviewers independently searched databases, selected studies to be included in the review and extracted data. If the two reviewers disagreed, the study was discussed in detail until they reached a consensus. Data for each study included were exported into a Microsoft Office Excel 2007 database file. To provide a standardized format, units of measurement were recalculated to percentage contribution of LCPUFA to total fatty acid composition of the relevant biomarker (% weight/weight)

from original data in the publication. When it was not possible to convert data, we contacted the authors for details.

4.2.4. Statistical analysis

Statistical analyses were performed using the Review Manager 5.1 Software (The Cochrane Collaboration, Oxford, United Kingdom), with the random-effects model. The confidence interval (CI) was established at 95%. P values of less than 0.05 were considered to indicate statistical significance. Statistical heterogeneity was assessed using the I² statistics (I² of 50% or more indicating the presence of significant heterogeneity).

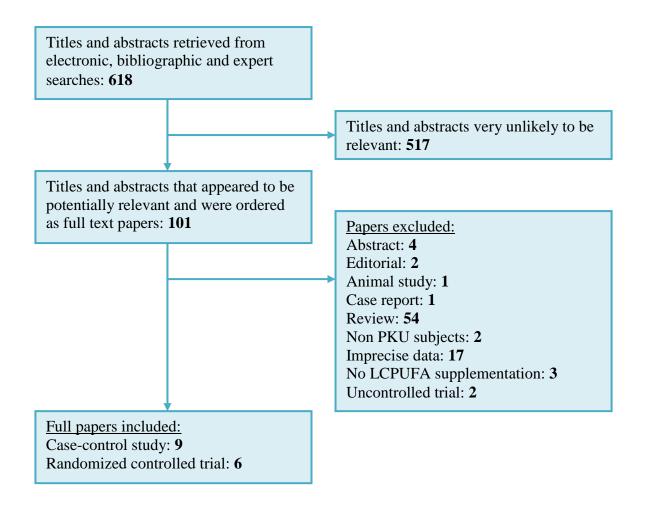


Figure 8. Flow diagram of the systematic literature search

4.2.5. Results

The flow diagram of the literature search for this review is shown in **Figure 8**. We included nine case-control studies and six RCTs in this systematic review, with a total of 713 and 273 participants, respectively.

Table 5. Basic characteristics of included case-control studies

Study	Country	Age	No. of PKU	No. of healthy	Biomarkers
			patients	controls	reported
Acosta et al	USA	1 – 13 yr	28	26	P, E
2001					
Galli et al 1991	Italy	3 – 12 yr	15	12	P, PPL, PCE, E
Giovannini et al	Italy	9 – 14 yr	45	45	PPL
2011					
Lage et al 2010	Spain	6 – 42 yr	47	77	P; PPL
Moseley et al	USA	10 – 50 yr	27	120	P, E
2002					
Sanjurjo et al	Spain	2 mo – 20 yr	40	50	P, EPL
1994					
Yi et al 2011a	USA	>12 yr	41	25	P, E
		-			
van Gool et al	the	6 mo – 25 yr	9	18	PPL, EPL
2000	Netherlands	•			

Abbreviations: P, total plasma; E, erythrocyte membrane; PKU, phenylketonuria, PPL, plasma phospholipids; PCE, plasma cholesteryl esters; EPL, erythrocyte phospholipids.

4.2.5.1. Case-control studies investigating fatty acid compositional differences between PKU patients and healthy controls

In the case-control studies included, fatty acid composition of the following 5 biomarkers was reported: total plasma lipids, plasma phospholipids, plasma cholesteryl esters, total erythrocyte membrane lipids and erythrocyte phospholipids. Basic characteristics of the studies included are presented in **Table 5**. In three papers the diagnosis of participants was classic PKU (Acosta et al. 2001; Sanjurjo et al. 1994; van Gool et al. 2000), two studies investigated also patients with moderate or mild PKU subtypes (Phe levels lower than 1200 or 600 μ mol/L) (Lage et al. 2010; Vilaseca

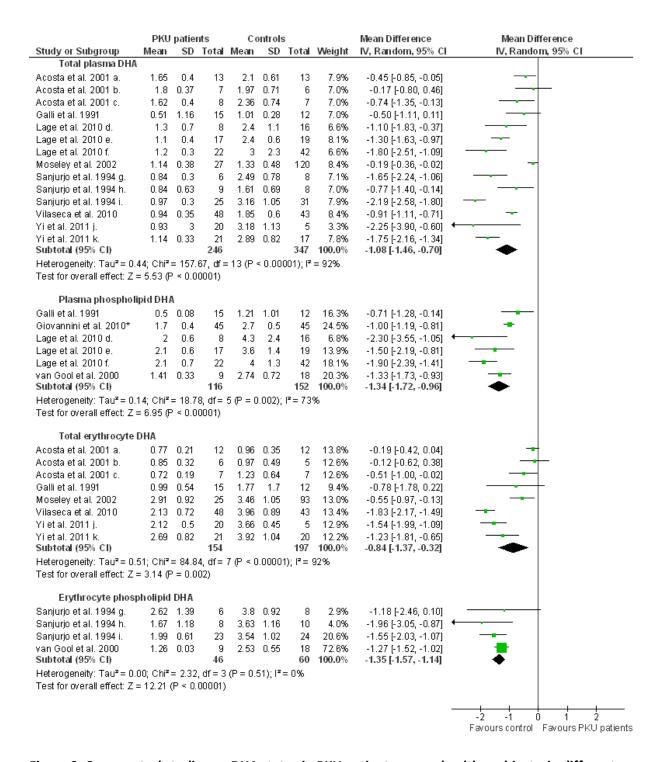


Figure 9. Case-control studies on DHA status in PKU patients versus healthy subjects, in different biomarkers

et al. 2010), whereas in four papers no further subclassification of PKU was described (Galli et al. 1991; Giovannini et al. 2011; Moseley et al. 2002; Yi et al. 2011). All patients followed amino acid-modified diets. Bad metabolic compliance of participants (Phe values higher than 600 μ mol/L, in spite of dietary intervention) was reported in two of the studies included (Moseley et al. 2002; Yi et al. 2011).

In this review we focused on linoleic acid (LA) and arachidonic acid (AA) from the n-6 series and alpha-linolenic acid (ALA), eicosapentaenoic acid (EPA) and docosahexaenoic acid (DHA) from the n-3 series. We conducted meta-analysis in cases of fatty acids and biomarkers used in at least three independent studies.

Fatty acid composition of total plasma lipids

Seven papers reported fatty acid composition of plasma total lipids. Analysis showed significantly lower contribution of the n-6 LCPUFA, AA (MD: -0.75; 95% CI: -1.31, -0.18; 12 studies; 596 participants; $I^2 = 68\%$) and the n-3 LCFUFA, EPA (MD: -0.20; 95% CI: -0.27, -0.13; 11 studies; 505 participants; $I^2 = 77\%$) and DHA (for data see **Figure 9**) to plasma total lipids of patients with PKU compared to healthy controls, while in the values of the essential precursors, LA (MD: 1.27; 95% CI: -0.23, 2.76; 12 studies; 596 participants; $I^2 = 65\%$) and ALA (MD: 0.05; 95% CI: 0.00, 0.10; 11 studies; 569 participants; $I^2 = 73\%$) there was no significant difference between the two groups.

Fatty acid composition of plasma phospholipids

Four papers reported fatty acid composition of plasma phospholipids. Primary analysis showed significantly lower EPA (MD: -0.41; 95% CI: -0.68, -0.14; 6 studies; 268 participants; $I^2 = 94\%$) and DHA (**Figure 9**) values in patients with PKU than in controls. No significant differences were found between the two groups with respect to AA (MD: 0.00; 95% CI: -0.80, 0.81; 6 studies; 268 participants; $I^2 = 30\%$), LA (MD: 2.62; 95% CI: -0.46, 5.71; 6 studies; 268 participants; $I^2 = 88\%$) or ALA values (MD: -0.07; 95% CI: -0.17, 0.03; 5 studies; 241 participants; $I^2 = 97\%$).

Fatty acid composition of erythrocyte membrane total lipids

Comparison of erythrocyte membrane total lipid fatty acid composition in patients with PKU and healthy controls was present in 8 studies. The primary analysis showed significantly lower EPA (MD: -0.08; 95% CI: -0.16, -0.01; 7 studies; 260 participants; $I^2 = 73\%$) and DHA (**Figure 9**) values in patients as compared to controls; however, there was no significant difference in AA values (MD: -0.32; 95% CI: -1.07, 0.43; 8 studies; 351 participants; $I^2 = 71\%$). There was no difference in LA values between the two groups (MD: 0.53; 95% CI: -0.18, 1.23; 8 studies; 351 participants; $I^2 = 52\%$), whereas ALA values were significantly higher in patients than in controls (MD: 0.03; 95% CI: 0.01, 0.05; 7 studies; 324 participants; $I^2 = 43\%$).

Fatty acid composition of erythrocyte phospholipids

Data relating to the erythrocyte phospholipid fraction were sufficient only for a metaanalysis of DHA; values were significantly lower in PKU patients than in controls (**Fig-ure 9**).

Fatty acid composition of plasma cholesteryl esters

There were insufficient data to conduct a meta-analysis comparing fatty acid composition of plasma cholesteryl esters in subjects with PKU and controls.

4.2.5.2. Effects of LCPUFA supplementation on DHA concentrations in PKU

We included six RCTs with parallel design. In two studies participants were newborns (Agostoni et al. 2006; Koletzko et al. 2007), in three studies infants and children between 1 and 18 years (Agostoni et al. 2003; Agostoni et al. 1995; Cleary et al. 2006) were included, while one study included both children and adults (Yi et al. 2011). Five studies were carried out in Europe, one in the USA (Yi et al. 2011).

In this report we primarily focus on the effect of LCPUFA supplementation on DHA status. We found eight biomarkers used to characterize changes in DHA values. From these, only total plasma DHA was used in at least three independent studies; never-

theless, the effect of LCPUFA supplementation on every biomarker of DHA status is shown in **Figure 10**. Agostoni et al (1995) applied supplementation with 2.5–4 g fish oil (18 g EPA, 4 g DPA and 12 g DHA/100 g fatty acid) daily for 6 months; Agostoni et al (2003) used 1 capsule (37 mg AA, 27.5 mg EPA, 20 mg DPA and 40 mg DHA/0.5 g capsule) per 4 kg body weight for 1 year; Yi et al (2011) used microalgae oil capsules (10 mg/kg/day DHA) for 4.5 months of supplementation; Koletzko et al (2007) used a supplemented formula (0.46 g AA and 0.27 g DHA/100 g fatty acids) for 1 year; Agostoni et al (2006) used a supplemented formula with slightly different composition (0.7 g AA and 0.3 g DHA/100 g fatty acids) for 1 year while Cleary et al (2006) gave essential fatty acid supplemented protein substitute (17.2 g LA and 4.5 g ALA/100 g fatty acid) for 20 weeks. Although different dosages and forms of n-3 LCPUFA supplementation were used in the different studies, these were all effective in significantly increasing DHA values of different biomarkers (**Figure 10**).

	Suppleme	_	•		rolgro	•	Mean Difference	Mean Difference
Study or Subgroup	Mean	SD	Total	Mean	SD	Total	IV, Random, 95% CI	IV, Random, 95% CI
1.2.1 Total plasma DHA								
Agostoni et al. 1995 A.	2.94	0.88	10	0.73	0.08	11	2.21 [1.66, 2.76]	-
Agostoni et al. 2003 B.	2.3	1.1	10	1.1	0.3	10	1.20 [0.49, 1.91]	—
Yi et al. (II.) 2011 C.	3.14	0.57	7	0.97	0.34	11	2.17 [1.70, 2.64]	-
1.2.2 Plasma phospholig	oid DHA							
Agostoni et al. 2003 B.	3.1	1.6	10	1.6	0.4	10	1.50 [0.48, 2.52]	- -
Koletzo et al. 2007 D.	3.08	0.32	10	1.52	0.63	11	1.56 [1.14, 1.98]	
Noiemo et al. 2007 D.	3.00	0.32	10	1.52	0.03		1.50 [1.14, 1.50]	
1.2.3 Plasma cholestery	l ester DHA							
Agostoni et al. 2003 B.	0.5	0.2	10	0.2	0.1	10	0.30 [0.16, 0.44]	+
1.2.4 Plasma triacylgyce								
Agostoni et al. 2003 B.	0.6	0.5	10	0.3	0.3	10	0.30 [-0.06, 0.66]	
1.2.4 Total erythrocyte D Agostoni et al. 2003 B.	2.8	1.5	10	1.5	0.5	10	1.30 [0.32, 2.28]	
Yi et al. (II.) 2011 C.	5.82	1.26	7	2.35	0.78	11	3.47 [2.43, 4.51]	
1.2.5 Erythrocyte phospi	•							
Agostoni et al. 2006 E.	3.6	1.06	17	1.4	0.44	11	2.20 [1.63, 2.77]	
Cleary et al. 2006 F.	2.07	0.8	19	1.64	0.4	11	0.43 [-0.00, 0.86]	
1.2.6 Erythrocyte phospi	hatidylcholi	ine DHA						
Agostoni et al. 2003 B.	0.8	0.2	10	0.3	0.2	10	0.50 [0.32, 0.68]	+
1.2.7 Erythrocyte phospi	-							
Agostoni et al. 2003 B.	3.7	1.7	10	1.3	0.9	10	2.40 [1.21, 3.59]	- + -
								-2 -1 0 1 2 Intervention lower Intervention higher

Figure 10. Randomized controlled trials on the effect of LCPUFA supplementation on DHA status in PKU

4.2.6. Discussion

The mainstream of phenylketonuria management is the restriction of dietary intake of phenylalanine, immediately after confirmation of the diagnosis. This means that patients with phenylketonuria have to consume a phenylalanine free formula and avoid foods rich in protein (e.g. meat, fish, egg, cheeses, normal bread and seeds), moreover, they have to avoid foods and drinks containing aspartame, flour, soya, beer or cream liqueurs. Low protein natural foods (potatoes, some vegetables and most cereals) can be consumed in restricted amounts (Blau et al. 2010). This strict diet may lead to the insufficient intake of some essential nutrients.

The potentially inadequate dietary LCPUFA supply of patients suffering from PKU on Phe-restricted diet has gained attention recently. There are two main sources of LCPUFA in the human organism: dietary intake and endogenous synthesis from their essential metabolites, LA and ALA (**Figure 1**). However, the endogenous synthesis of AA and DHA is limited (Calder 2012), therefore, the lack of preformed LCPUFA in the diet may lead to deficiency. Moreover, in PKU a possible inhibitory effect of Phe metabolites on endogenous DHA synthesis is suspected (Infante and Huszagh 2001).

The connection between elevated plasma Phe levels and neurological disorders seen in patients with PKU is well-established (MacDonald et al. 2011). The optimum outcome is mainly dependent on metabolic control with diet and this control varies during the patient's life. However, functional deficits may also be present in PKU patients treated early and well. Patients with PKU have lower intelligence quotients as compared to healthy controls (Brumm and Grant 2010). Subtle abnormalities in phenylketonuria include the impairment of executive abilities (such as planning, problem solving, information processing and sustained attention (Van Zutphen et al. 2007)). Besides, children with phenylketonuria have behavioral abnormalities, motor dysfunction and impaired memory (Blau et al. 2010).

LCPUFA are shown to have important role in cognitive development, in maturation of visual acuity and development of motor functions in full-term and preterm infants (Fleith and Clandinin 2005). On the basis of these considerations it has been hypothesized that LCPUFA deficiency, especially inadequate DHA supply, might contribute also to the neurological abnormalities observed in patients with PKU (Koletzko et al. 2009).

In this review we included 9 case-control studies (divided into 16 arms) and six RCTs (divided into 13 arms) in order to assess whether significant deficiency of LCPUFA can be demonstrated in patients with PKU. We also investigated whether LCPUFA status of PKU patients can be improved by supplementing their diet with LCPUFA. We found that in PKU patients on low-protein diet blood levels of the two principal n-3 LCPUFA, EPA and DHA, were consequently and significantly reduced in different blood biomarkers; whereas the values of the principal n-6 LCPUFA, AA were significantly reduced only in plasma total lipids. These data indicate that supplementation with oils containing DHA may be an effective way for improving the n-3 LCPUFA status of patients with PKU. Special dietary products for infants with PKU are usually enriched with n-3 and n-6 LCPUFA, however, supplementation should be applied routinely also after infancy. This may be achieved relatively easily since large numbers of various capsules containing fish oil are available.

A limitation of the present review was that in the different studies different dosages of LCPUFA were used for supplementation, and therefore subgroup analysis according to supplement dosage was not possible. A further limitation was the lack of description of compliance and blood Phe levels of the patients with PKU on low-protein diet; therefore, subgroup analysis evaluating the potential influence of different blood Phe levels on PUFA metabolism was not possible. We assume that insufficient LCPUFA intakes together with metabolic impairment of LCPUFA synthesis may lead to decreased LCPUFA levels seen in patients with PKU.

The data systematically reviewed here indicate that in patients with PKU n-3 LCPUFA supply is insufficient. However, data from RCTs suggest that DHA status in patients with PKU may be improved by dietary supplementation. Further studies are needed to investigate whether LCPUFA deficiency is present in all age groups, especially in adolescents and adults who usually follow more relaxed diets. Moreover, there is only limited evidence about the optimal LCPUFA supplementation dosage in the different age groups.

4.3 Prebiotics in healthy infants and children for prevention of acute infectious diseases: a systematic review and meta-analysis

4.3.1. Inclusion criteria

To be included, a study needed to meet all of the following criteria: a) randomized allocation to treatment groups (RCT); b) carried out in healthy infants or children, aged 0-18 years; c) intervention with prebiotics (prebiotics added to food in the manufacturing process or as a separate supplement) compared to controls (placebo or no supplementation); d) a supplementation time of at least two months and an observation time of at least four months; e) using one of the following supplements: oligosaccharides, galacto-oligosaccharides (GOS), fructo-oligosaccharides (FOS), fructans, inulin, oligofructose; f) describing at least one of the following outcomes: incidence of overall infections (any kind of infections), incidence of all acute respiratory tract infections (RTI), incidence of upper respiratory tract infections (URTI), incidence of lower respiratory tract infections (LRTI), incidence of otitis media, incidence of gastrointestinal infections (GITI) (defined in primary studies as 3 or more liquid/semiliquid stools per day with fever, vomiting, and/or dehydration and compromised general status), incidence of diarrhea episodes, incidence of urinary tract infections (UTI), incidence of fever episodes, incidence of infections requiring antibiotic treatment. No language restrictions were applied.

4.3.2. Search Strategy

Ovid MEDLINE (www.ovid.com), Scopus (www.scopus.elsevier.com), Web of Science (www.webofknowledge.com) and the Cochrane Library CENTRAL database (www.thecochranelibrary.org) were searched from inception to Week 3, June,2013 for intervention studies using text terms with appropriate truncation and relevant indexing terms. The search was in the form: [prebiotics terms] and [search filters for children] and [search filters for randomized controlled trials]. For the Ovid MEDLINE

search we used the sensitivity- and precision-maximizing version (2008 revision) of the Cochrane Highly Sensitive Search Strategy developed for identifying randomized trials in MEDLINE (Lefebvre et al. 2011). The full search strategy for the Ovid MEDLINE database can be found in **Table 6**, search of the other databases was also based on this strategy.

Table 6. Search strategy for Ovid MEDLINE 1946 to June Week 3 2013

# Search History	Results
1 exp Prebiotics/	632
2 prebiotic*.mp.	3213
3 exp Oligosaccharides/	71620
4 oligosaccharid*.mp.	34893
5 oligo-saccharid*.mp.	90
6 fructooligosaccharid*.mp.	490
7 galactooligosaccharid*.mp.	202
8 exp Fructans/	6464
9 (fructan or fructans).mp.	1016
10 (inulin or inulins).mp.	9338
11 (oligofructos* or oligo-fructos*).mp.	343
12 2 or 3 or 4 or 5 or 6 or 7 or 8 or 9 or 10 or 11 or 12	96524
13 limit 13 to "all child (0 to 18 years)"	6010
(child* or boy* or girl* or adolescent* or pediatric* or paediatric* or infant* or newborn* or neonat* or toddle* or schoolchild*).mp.	3190789
15 13 and 15	7476
16 14 or 16	7476
17 randomized controlled trial.pt.	367158
18 controlled clinical trial.pt.	87691
19 randomi#ed.ab.	318588
20 placebo.ab.	144978
21 clinical trials as topic.sh.	169978
22 randomly.ab.	187790
23 trial.ti.	115019
24 18 or 19 or 20 or 21 or 22 or 23 or 24	860275
25 exp animals/ not humans.sh.	3910958
26 25 not 26	792849
27 17 and 27	1072

4.3.3. Data extraction and management

Titles and abstracts were screened for inclusion by a single reviewer. All potentially relevant abstracts and full papers were screened for inclusion using an inclusion/exclusion form by two reviewers independently; where they disagreed the study was discussed with a third reviewer. Data for each included study were extracted by two reviewers independently. When it was necessary, units of measurement were converted to a standard form (number of episodes/person/year) to facilitate comparison across studies.

Assessment of risk of bias in the studies included

Two reviewers independently assessed the methodological quality of studies included. "Low risk of bias" meant that the study was randomized, the randomization method and allocation concealment was described, the study was double-blind, the method of blinding was described and the reasons for and numbers of dropouts were stated (or there were no dropouts) and the distribution of dropouts across comparison groups was equal. All other studies were considered as "moderate or high risk of bias".

4.3.4 Statistical analysis

Meta-analysis was carried out according to the methodology for counts and rates described in The Cochrane Handbook (Deeks and Altman 2011). The results of the studies were expressed as a rate ratio, the ratio of the rate in the prebiotic group to the rate in the control group.

$$rate \ ratio = \quad \frac{E_{prebiotic}/T_{prebiotic}}{E_{control}/T_{control}}$$

where $E_{prebiotic}$ means the events occurred during $T_{prebiotic}$ participant-years of follow-up in the prebiotic intervention group, and $E_{control}$ means the events during $T_{control}$ participant-years in the placebo group. The (natural) logarithm of the rate ratios was

combined across studies using the generic inverse-variance method. The standard error of the log rate ratio was expressed by

$$SE = \sqrt{\frac{1}{E_{prebiotic}} + \frac{1}{E_{control}}}$$

Assessment of heterogeneity

We carried out tests for heterogeneity using the Chi^2 test with significance being set at a p value < 0.1. We used the I^2 statistic to estimate the variation across studies. An I^2 statistic < 40% was considered to be a low level of heterogeneity, 30% to 60% a moderate level, 50% to 90 % a substantial level and 75% to 100 % a considerable level (Deeks and Altman 2011).

Data synthesis

Regardless of heterogeneity between the pooled studies, we used a random-effects model to synthesize all data.

4.3.5. Results

Study inclusion

The flow diagram of the literature search for this review is shown in **Figure 10**. Altogether 1429 titles and abstracts were identified via the electronic search, 39 of them appeared to be potentially relevant. One conference abstract could not be retrieved; all others were available for detailed assessment. Finally, 5 studies reported in 8 full-text papers and 4 conference abstracts fulfilled the inclusion criteria.

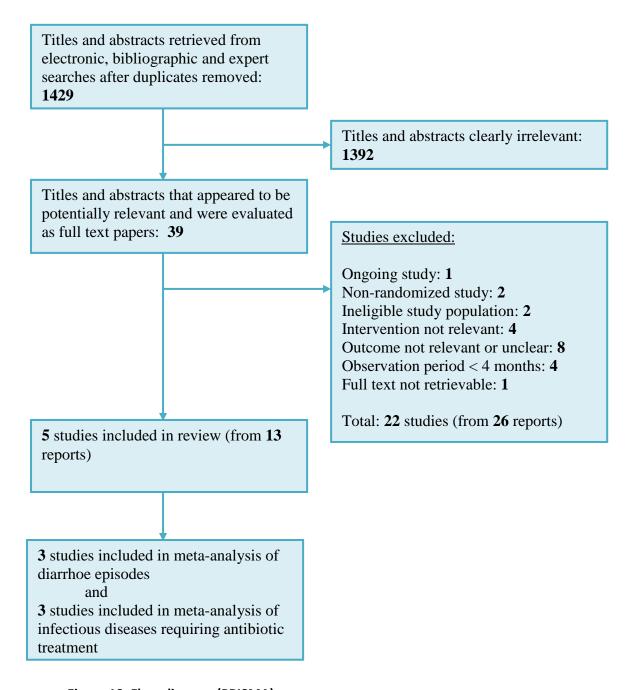


Figure 10. Flow diagram (PRISMA)

The main characteristics of the 5 included studies are described in **Table 7**.

Participants

The studies ranged in size from 140 to 830 children randomized initially. All participants were infants and children younger than two years of age at the beginning of the intervention. The countries where trials were performed are also listed in **Table 7**.

Interventions

The studies involved different types of prebiotics including GOS/FOS mixture (Arslanoglu et al. 2007, Bruzzese et al. 2009, Stuijvenberg et al. 2011), polydextrose (PDX)/GOS mixture (Ribeiro et al. 2012) or oligofructose (Duggan et al. 2003, Saavedra and Tschernia 2002). In most of the studies included the intervention and observation time was the same and ranged between 108 days and 12 months, there was only one study (Arslanoglu et al. 2008, Arslanoglu et al. 2007) in which investigators kept on observing the patients after the end of supplementation (in this study participants were followed for two years). To ensure homogenous data we included incidence data into the meta-analysis from the study mentioned above, starting from the time point the supplementation was finished.

Risk of bias in the studies included

Aspects of methodological quality of the studies included are described in **Table 8**. In two of the five studies included (Arslanoglu et al. 2008, Duggan et al. 2003, Stuijvenberg et al. 2011) method of randomization, methods of blinding and reasons for dropouts were all properly described, which did not suggest high risk of bias.

Table 7. Characteristics of included studies

Author, publication year		Participants		Intervention	Outcome	
cation year	Country	Inclusion criteria	Supplement used	Placebo used	Duration of sup- plementation	Relevant outcomes data available for ¹
Arslanoglu et al. (2008/2007) (34, 35)	Italy	Healthy term infants, start of formula feeding within the first two weeks of life, parental history of atopic disease	Hypoallergenic formula supple- mented with 0.8 g/100 ml scGOS/IcFOS	Hypoallergenic formula sup- plemented with 0.8 g/100 ml maltodextrin	6 months	Incidence of physician-diagnosed infections (URTI, LRTI, otitis media, GITI, UTI), Antibiotic prescriptions, Incidence of fever episodes witnessed by the parents
Bruzzese et al. (2009) (37)	Italy	Healthy term infants, aged 15-120 days old, birth weight >2500g, start of formula feeding after at least 15 days of exclusive breastfeeding	Standard formula supplemented with 0.4 g/100 ml GOS/FOS (9:1)	Standard infant formula	12 months	Incidence of acute diarrhea, Incidence of URTI, Incidence of LRTI, Incidence of respiratory infections requiring AB therapy
Duggan et al. (2003) (38)	Peru	Healthy infants, aged 6-12 months, already consuming solid food	Rice- or oat-based cereal supple- mented with 0.55 g oligofructose /15 g cereal	Non- supplemented cereal	6 months	Incidence of episodes of diarrhea

Saavedra et al. (2002) (40)	USA	Healthy infants and children, aged 4-24 months; already consuming cereals, attending daycare	cereal supple- mented with 0.55 g oligofructose /15 g cereal (administered ad libitum with a min- imum goal intake of 15 g cereal/day)	Non- supplemented cereal	6 months	Incidence of diarrhea by parental report, Incidence of diarrhea accompanied by fever and vomiting, Incidence of antibiotic use
Stuijvenberg et al. (2011) (43)	The Netherlan ds, Austria, Switzerla nd, Italy, Germany	Healthy term infants, aged < 8 weeks; birth weight above the 10 th percentile for gestational age, without parental history of atopic disease	Standard formula supplemented with 0.8 g/100 ml GOS/FOS (9:1)	Standard infant formula	10-12 months	Incidence of fever episodes witnessed by the parents, Incidence of AB treatments reported by the parents

¹Total number of episodes was converted into incidence of episodes, where necessary

Abbreviations: GOS, galacto-oligosaccharides; FOS, fructo-oligosaccharides; scGOS, short chain GOS; lcFOS, long chain FOS; PDX, polydextrose; URTI, upper respiratory tract infection; LRTI, lower respiratory tract infection; RTI, respiratory tract infection; GITI,

Table 8. Methodological quality of included studies (Risk of bias)

	Selection bias			Performan	ce/Detection bias		Attrition bias			Overall risk of bias ¹
Trial	Study design (rando- mized: YES/NO/u nclear)	Method of ran- domization (clearly described and appropriate: YES/NO)	Allocation concealment (clearly described and appropriate: YES/NO)	Blinding (double- blind: YES/NO/ unclear)	Methods of blinding appropriate (YES/NO/unclear)	Description of withdrawal or dropout (YES/NO)	Reason for drop- outs in the inter- vention group (Number of drop- outs)	Distribution of dropouts across comparison groups (Num- ber of drop- outs/Total included)	All out- comes listed in Methods described in Results (YES/NO/u nclear)	
Arslanoglu et al. (2008/2007)	YES	YES "a preprepared randomization numbers table"	NO	YES	unclear	YES	- In the first 6 months: Reestablishment of breastfeeding (22), Move to another city (8/), Flatulence, no reason (5) - In the 6-24 months follow up period: Poor compliance (8)	Intervention (I): 27/129 Control (C): 26/104	YES	Moderate/ High
Bruzzese et al. (2009)	YES	YES "a random numbers table with a block design"	NO	NO "open trial"	NO	YES	Poor compliance (30), Development of cow's milk protein intolerance (3), Excluded (incomplete data) (40)	I: 73/169 C: 68/173	YES	Moderate/ High
Duggan et al. (2003)	YES	YES "random assignments (using a permuted block	YES "on enrollment of an infant, study staff opened the	YES	YES "personnel not involved in the study generated	YES	Moved (6), Refused blood sample (4), Ill with non-diarrhea	I: 12/141 C: 19/141	YES	Low

		design"	next study envelope		the list of random		illness (1),			
			to determine which		assignments and		Withdrew consent			
			cereal was to be		prepared enve-		(1)			
			dispensed to that		lopes containing					
			infant"		the cereal as-					
					signments"					
Saavedra et	YES	NO	NO	YES	unclear	YES	No dropouts	I: 0	unclear	Moderate/
al. (2002)								C: 0		High
Stuijvenberg	YES	YES	YES	YES	YES	YES	Change to another	I: 53/414	YES	Low
et al. (2011)		"Time-balanced	"only the hospital		"The parents, the		formula (8),	C: 42/416		
		randomization	pharmacist had a		study physicians		Intolerance to			
		was performed	copy of the ran-		and the study		formula (10),			
		with the software	domization list with		nurses were		Occurrence of any			
		RANCODE, with a	the actual treat-		unaware of the		disease (7),			
		random permuted	ment allocation"		group allocation"		Withdrawal (28)			
		block size of 4"								

¹ "Low risk of bias" meant that the study was randomized, the randomization method and allocation concealment was described, the study was double-blind, the method of blinding was described, the reasons for and numbers of dropouts were stated (or there were no dropouts) and the distribution of dropouts across comparison groups was equal. All other studies were considered as "moderate or high risk of bias".

Patient-relevant outcomes described in the studies included

Incidence of overall infection

The incidence of any infection during the observation period was reported only in one study (35): infants in the scGOS/IcFOS group had significantly fewer episodes of any type of infection during the 6-month supplementation-observation period (0.42 vs. 0.90 episodes/subject/year; $n_{prebiotic} = 102$, $n_{control} = 104$; p = 0.01; RR 0.47, 95% CI 0.32 – 0.68).

Incidence of fever episodes

The incidence of fever episodes recorded by the parents of the participating infants and children up to 24 months was examined in two studies. Saavedra et al (2002) observed a statistically significant difference in the incidence of fever with 'cold symptoms' between the intervention and the control groups (5.76 vs. 9.05 fever episodes/subject/year; $n_{prebiotic} = 63$, $n_{control} = 60$; p < 0.05). However, Stuijvenberg et al (2011) found no statistically significant difference in the number of fever episodes during the one-year supplementation period between the prebiotic and control group (1.79 vs. 1.78 fever episodes/subject/year; $n_{prebiotic} = 292$, $n_{control} = 300$; p > 0.05). The pooled effect of these two studies was not significant either (RR 0.80, 95% CI 0.51 – 1.26) (**Figure 11**) but showed a considerable level of heterogeneity (Chi² 24.89; df = 1, P < 0.00001; I² statistic = 96%).

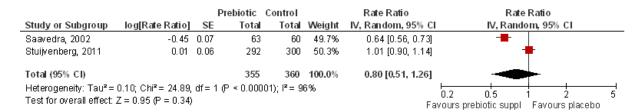


Figure 11. The rate of fever episodes (events per person/year) in infants and children supplemented with prebiotics versus placebo

Incidence of infections requiring antibiotic therapy

For three RCTs eligible count data were available on the effect of prebiotic supplementation on the incidence of infections requiring antibiotic treatment. A pooled analysis of the data from these 3 studies showed a statistically significant difference in the episode rates of acute bacterial infections (RR 0.68, 95% CI 0.61 - 0.77) (**Figure 12**). The level of heterogeneity between these studies was low (Chi² 1.04; df = 2, P = 0.59; I² statistic = 0%).

Stuijvenberg et al (2011) reported their data on fever episodes requiring antibiotics as median (25th–75th percentile) and so this study was not included in the meta-analysis. It has to be noted, though, that in this study no significant difference in the median adjusted numbers of fever episodes requiring antibiotics was described (prebiotic group: 0.05 (0.05, 0.11) vs. control group: 0.05 (0.05, 0.16), median values (25th, 75th percentile)).

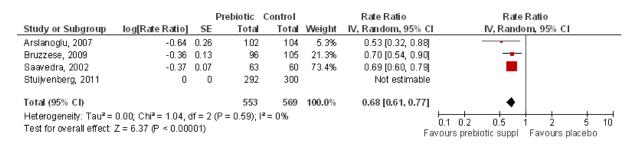


Figure 12. The rate of infections requiring antibiotic treatment (events per person/year) in infants and children supplemented with prebiotics versus placebo

Incidence of gastrointestinal infections or diarrhea

Count data on any kind of gastrointestinal infections (GITI) were available for one RCT (35); the rate of GITI was lower in the prebiotic group, however this difference was not statistically significant (0.02 vs. 0.08 episodes/subject/year; p = 0.18). One further study (Bruzzese et al. 2009) only reported that the mean number of episodes of gastroenteritis was significantly lower in the GOS/FOS group than in controls (p = 0.01).

Diarrhea episodes were reported for three RCTs (Bruzzese et al. 2009, Duggan et al. 2003, Saavedra and Tschernia 2002). The pooled effect estimate of these three studies shows no significant difference (RR 0.71, 95% CI 0.35 – 1.43) between the prebiotic supplemented and the placebo groups on the rate of diarrhea episodes (**Figure 13**), however, the level of heterogeneity among these studies was considerable (Chi² 356.65; df = 2, P < 0.00001; I^2 statistic = 99%).

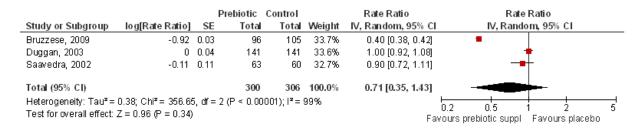


Figure 13 The rate of diarrhoea episodes (events per person/year) in infants and children supplemented with prebiotics versus placebo

Incidence of respiratory tract infections

Information on the incidence of upper respiratory tract infections (URTI) was available for 2 RCTs. Both studies (Arslanoglu et al. 2007, Bruzzese et al. 2009) only reported that the number of episodes of URTI was lower in the prebiotic group as compared to the control group, but the difference was in neither of the two studies significant. Arslanoglu et al (2007) observed no significant difference between the prebiotic supplemented and the control group either in the rate of otitis media (0.08 vs. 0.12 episodes/subject/year; p = 0.60) or in the rate of lower respiratory tract infections (0.04 vs. 0.06 episodes/subject/year; p = 0.07).

Incidence of urinary tract infections

One report (Arslanoglu et al. 2007) provided data on the effect of prebiotic supplementation on the incidence of urinary tract infections; it showed no significant difference in the incidence between the two groups (0.04 vs. 0.14 episodes/subject/year; p = 0.26).

4.3.6. Discussion

This review was meant to investigate whether there is evidence for prebiotic oligo-saccharides to have a protective effect against acute infectious diseases in infancy, and furthermore, whether or not such a protective effect is present later in child-hood. Five studies investigated the relevant outcomes; however, there were only two outcomes (incidence of infections requiring antibiotic treatment and incidence of diarrhea episodes) for which eligible count data were available from three RCTs. For each further included outcome data were extracted from only one or two studies.

There is strong evidence (large sample size, low level of heterogeneity, moderate risk of bias) only for bacterial infections to have a significantly lower incidence in 0-24 month old infants and children regularly consuming prebiotics as compared to children supplemented with placebo.

Our review focused on the infection-preventive effect of prebiotics in the pediatric population; however, studies investigating the preventive efficacy of prebiotics in children aged 2-18 years are completely lacking in the literature. Based on the promising results seen in the 0-2 year-old age group it would be worth conducting such studies also in older children, especially because children newly entering a community (day nurseries, kindergartens, elementary schools) are acutely exposed to infections. Finding an effective strategy for the prevention of common infections would also decrease health expenses and thereby not only benefits for the individual but also for the entire society.

There were only two studies in which risk of bias could be labeled as low. The lack of describing allocation concealment and the method of blinding in the other three studies could potentially have biased the results favoring the prebiotic group (**Table 8**).

Most outcome measures in this review come from only one trial, therefore some effects of prebiotics may have remained hidden because of the small sample sizes for some outcomes. Future RCTs should consider the assessment of incidence data in

reference to overall infections, fever episodes, upper respiratory tract infections and urinary tract infections.

From the clinical point of view the relevant five studies included were homogenous: they all investigated infants or children aged 0-24 months, the supplementation was conducted for 6-12 months and incidence data were determined by the end of intervention.

However, when pooled analysis was performed for the outcome "Incidence of diarrhea episodes" high levels of heterogeneity was found. This may be a consequence of the different types of prebiotics used: the FOS/GOS mixture (Bruzzese 2009) seems to be more effective in the prevention of diarrhea episodes compared to oligofructose (Duggan et al. 2003, Saavedra et al. 2002). Another explanation for the heterogeneity may be the difference in the way of prebiotic administration: while in the study of Bruzzese et al the GOS/FOS mixture was added to the formula, in the other two studies cereals supplemented with oligofructose were used.

There were large differences in the baseline risk for infections among different populations: for example, the mean number of episodes/subject/year of bacterial infections was in the study of Saavedra et al (2002) 8 times higher than in the study of Bruzzese et al (2009), and 38 times higher than in the study of Arslanoglu et al (2007). Large differences were also seen in the mean number of diarrhea episodes/subject/year.

Finally, it has to be mentioned that Stuijvenberg et al (2011) reported their data on fever episodes requiring antibiotics as median values (25th–75th percentile) and so this study was not included in the meta-analysis. However, the rate of antibiotic use in this study was much lower than in the other three studies, indicating lower weight and therefore no determinative impact on the pooled effect estimate.

The role of prebiotics has been investigated most extensively in the field of infant nutrition (Boehm et al. 2003, Thomas et al. 2010). Oligosaccharides were demonstrated to represent the most important dietary factor in human milk, promoting the

development of beneficial intestinal flora (Bertino et al. 2012, Barile and Rastall 2013) and being the bioactive compound responsible for the positive health effects of human milk: HMOs were described to have a protective effect against gastrointestinal diseases and diarrhea (Salone et al. 2013, Lamberti et al. 2011) and against respiratory tract infections (Duijts et al. 2009, Bachrach et al. 2003, Chantry et al. 2006).

However, it has to be mentioned that because of their complex structure, oligosaccharides with a structure identical to that of HMOs are not yet available for dietary supplementation purposes; at present galactooligosaccharides (GOS), fructooligosaccharides (FOS) and oligofrucose are considered as the most relevant prebiotic oligosaccharides used in supplementation studies. In this review we could not confirm all the preventive health effects ascribed to HMOs also for non-human milk oligosaccharides.

4. Novel findings and practical applications

- **1.** Our systematic review based on 51 publications showed significantly lower contribution of both arachidonic and docosahexaenoic acid to plasma total lipids and plasma phospholipids in men than in women; in erythrocyte membrane total lipids DHA values were significantly lower in men than in women.
- **2.** In supplementation studies reporting fatty acid composition in serum PL, serum total lipids or erythrocyte membrane lipids, gender distribution should be regarded as significant potential confounding variable.
- **3.** The data systematically reviewed here indicate that in patients with PKU n-3 LCPUFA supply is insufficient. However, data from RCTs suggest that DHA status in patients with PKU may be reflectively improved by dietary supplementation.
- **4.** There is only limited evidence about the optimal LCPUFA supplementation dosage in the different age groups. It is of clinical importance to investigate whether LCPUFA deficiency is present in all age groups, especially in adolescents and adults who usually follow more relaxed diets.
- **5.** Currently available evidence suggests that the preventive use of prebiotics decreases the rate number of infections requiring antibiotic therapy in infants and children aged 0-24 months. Data from only one randomized controlled study indicate that prebiotics may also be effective in decreasing the rate of overall infections.
- **6.** Further studies should be carried out in the 3-18 years old age group to answer the question whether prebiotics can be considered for the prevention of acute infectious diseases also in the older pediatric age group.

6. Acknowledgement

I would like to express my gratitude to Professor Tamás Decsi who woke my interest already as a medical studentin in paediatric nutrition and later, as a PhD student in evidence-based medicine and gave me his advice, support and intellectual help.

I am extremely grateful to the colleagues from the German Cochrane Centre, especially to Professor Gerd Antes, Jörg Meerpohl, Daniela Küllenberg and Anette Blümle, for their scientific support and their friendship.

I am grateful to all of my co-workers from the Department of Paediatrics and from other collaborating institutes, especially to Katalin Fekete, for their valuable contribution.

To everyone else who assisted my work scientifically or in other ways.

Finally, I would like to thank my husband and my children, László and Eszter for their love and continuous support and understanding.

7. List of references

Acosta PB, Yannicelli S, Singh R et al. Intake and blood levels of fatty acids in treated patients with phenylketonuria. J Pediatr Gastroenterol Nutr 2001;33:253-9.

Agostoni C, Braegger C, Decsi T et al. Supplementation of N-3 LCPUFA to the diet of children older than 2 years: a commentary by the ESPGHAN Committee on Nutrition. J Pediatr Gastroenterol Nutr 2011;53:2-10.

Agostoni C, Harvie A, McCulloch DL et al. A randomized trial of long-chain polyunsaturated fatty acid supplementation in infants with phenylketonuria. Dev Med Child Neurol 2006;48:207-12.

Agostoni C, Riva E, Biasucci G et al. The effects of n-3 and n-6 polyunsaturated fatty acids on plasma lipids and fatty acids of treated phenylketonuric children. Prostaglandins Leukot Essent Fatty Acids 1995;53:401-4.

Agostoni C, Verduci E, Massetto N, Radaelli G, Riva E, Giovannini M. Plasma long-chain polyunsaturated fatty acids and neurodevelopment through the first 12 months of life in phenylketonuria. Dev Med Child Neurol 2003;45:257-61.

Ando T, Yanagihashi T, Wakisaka I, Tomari T. Relationship between serum fatty acid composition and life style and/or health status in a rural population in Kagoshima (in Japanese). Nihon Koshu Eisei Zasshi 1990;37:752–60.

Antonini FM, Bucalossi A, Petruzzi E, Simoni R, Morini PL, D'Alessandro A. Fatty acid composition of adipose tissue in normal, atherosclerotic and diabetic subjects. Atherosclerosis 1970;11:279–89.

Arab L, Akbar J. Biomarkers and the measurement of fatty acids. Public Health Nutr 2002;5:865–71.

Araki K, Akiyam Y, Murayama H, Fukase O, Yamamoto A, Fujiwara T, Oshiba K, Kawaraya C, Otachi J, Masumoto K. Relationship between dietary intake and serum fatty acids compositions of the residents in Hyogo Prefecture (in Japanese). Nihon Koshu Eisei Zasshi 1990;37:620–8.

Arslanoglu S, Moro GE, Boehm G. Early supplementation of prebiotic oligosaccharides protects formula-fed infants against infections during the first 6 months of life. J Nutr. 2007;137:2420-4.

Arslanoglu S, Moro GE, Schmitt J, Tandoi L, Rizzardi S, Boehm G. Early dietary intervention with a mixture of prebiotic oligosaccharides reduces the incidence of allergic manifestations and infections during the first two years of life. J Nutr. 2008;138:1091-5.

Ashley C, Johnston WH, Harris CL, Stolz SI, Wampler JL, Berseth CL. Growth and tolerance of infants fed formula supplemented with polydextrose (PDX) and/or galactooligosaccharides (GOS): double-blind, randomized, controlled trial. Nutr J. 2012;11:38.

Bachrach VR, Schwarz E, Bachrach LR. Breastfeeding and the risk of hospitalization for respiratory disease in infancy: a meta-analysis. Arch Pediatr Adolesc Med. 2003;157:237-43.

Bakewell L, Burdge GC, Calder PC. Polyunsaturated fatty acid concentrations in young men and women consuming their habitual diets. Brit J Nutr 2006;96:93–9.

Barile D, Rastall RA. Human milk and related oligosaccharides as prebiotics. Current opinion in biotechnology. 2013;24:214-9.

Bertino E, Peila C, Giuliani F, Martano C, Cresi F, Di Nicola P, et al. Metabolism and biological functions of human milk oligosaccharides. J Biol Regul Homeost Agents. 2012;26:35-8.

Beynen AC, Hermus RJ, Hautvast JG. A mathematical relationship between the fatty acid composition of the diet and that of the adipose tissue in man. Am J Clin Nutr 1980;33:81–5.

Blau N, van Spronsen FJ, Levy HL. Phenylketonuria. Lancet 2010;376:1417-27.

Boehm G, Fanaro S, Jelinek J, Stahl B, Marini A. Prebiotic concept for infant nutrition. Acta Paediatr Suppl. 2003;91:64-7.

Boehm G, Jelinek J. Double-blind, controlled and randomised study with a parallel group design on the effect of formula feeding (IF and FOF) supplemented with a mixture of immunological active neutral and acidic oligosaccharides on the incidence of febrile respiratory and gastrointestinal infections in healthy term born infants during the first year of life. German Clinical Trials Register 2009.

Bolton-Smith C, Woodward M, Tavendale R. Evidence for age-related differences in the fatty acid composition of human adipose tissue, independent of diet. Eur J Clin Nutr 1997;51:619–24.

Bone A, Kuehl AK, Angelino AF. A neuropsychiatric perspective of phenylketonuria I: overview of phenylketonuria and its neuropsychiatric sequelae. Psychosomatics 2012; 53:517-23.

Bradbury J. Docosahexaenoic acid (DHA): an ancient nutrient for the modern human brain. Nutrients. 2011;3:529-54.

Brouwer DA, van der Dijs FP, Leerink CB, Steward HN, Kroon TA, Suverkropp GH, Romer JW, van Doormaal JJ, Muskiet FA. The dietary fatty acids of patients with coronary artery disease and controls in Curacao. Implications for primary and secondary prevention. West Indian Med J 1997;46:53–6.

Brumm VL, Grant ML. The role of intelligence in phenylketonuria: a review of research and management. Mol Genet Metab 2010;99:S18-21.

Bruzzese E, Volpicelli M, Squeglia V, Bruzzese D, Salvini F, Bisceglia M, et al. A formula containing galacto- and fructo-oligosaccharides prevents intestinal and extraintestinal infections: an observational study. Clin Nutr. 2009;28:156-61.

Burdge GC, Wootton SA. Conversion of alpha-linolenic acid to eicosapentaenoic, docosapentaenoic and docosahexaenoic acids in young women. Br J Nutr 2002;88:411– 20.

Calder PC. Mechanisms of action of (n-3) fatty acids. J Nutr 2012;142:592S-599S.

Campoy C, Escolano-Margarit MV, Anjos T, Szajewska H, Uauy R. Omega 3 fatty acids on child growth, visual acuity and neurodevelopment. Br J Nutr 2012;107:S85-S106.

Chantry CJ, Howard CR, Auinger P. Full breastfeeding duration and associated decrease in respiratory tract infection in US children. Pediatrics. 2006;117:425-32.

Chen H, Zhuo Q, Yuan W, Wang J, Wu T. Vitamin A for preventing acute lower respiratory tract infections in children up to seven years of age. Cochrane Database Syst Rev. 2008:Cd006090.

Cheng HH, Wen YY, Chen C. Serum fatty acid composition in primary school children is associated with serum cholesterol levels and dietary fat intake. Eur J Clin Nutr 2003;57:1613–20.

Childs CE, Roume-Nadal, Burdge, Calder PC. Gender differences in the n-3 fatty acid content of tissues. Proc Nutr Soc 2008;67:19–27.

Christensen JH, Christensen MS, Dyerberg J, Schmidt EB. Heart rate variability and fatty acid content of blood cell membranes: a dose-response study with n-3 fatty acids. Am J Clin Nutr 1999;70:331–7.

Cleary MA, Feillet F, White FJ et al. Randomised controlled trial of essential fatty acid supplementation in phenylketonuria. Eur J Clin Nutr 2006;60:915-20.

Conway SP, Phillips RR, Panday S. Admission to hospital with gastroenteritis. Arch Dis Child. 1990;65:579-84.

Dang D, Zhou W, Lun ZJ, Mu X, Wang DX, Wu H. Meta-analysis of probiotics and/or prebiotics for the prevention of eczema. The Journal of international medical research. 2013.

Decsi T, Kennedy K. Sex-specific differences in essential fatty acid metabolism. Am J Clin Nutr. 2011;94:1914S-9S

Decsi T, Koletzko B. Role of long-chain polyunsaturated fatty acids in early human neurodevelopment. Nutr Neurosci 2000;3:293-306.

Deeks JJ HJ, Altman DG (editors). Analysing data and undertaking meta-analyses. In: Higgins JPT GS, editor. Cochrane Handbook for Systematic Reviews of Interventions Version 510 (updated March 2011). www.cochrane-handbook.org: The Cochrane Collaboration; 2011.

Deeks JJ, Higgins JPT, Altman DG (editors). Chapter 9: Analysing data and undertaking meta-analyses. In: Higgins JPT, Green S (editors). Cochrane Handbook for Systematic Reviews of Interventions Version 5.1.0 (updated March 2011). The Cochrane Collaboration, 2011. Available from www.cochrane-handbook.org.

Del-Rio-Navarro BE, Espinosa Rosales F, Flenady V, Sienra-Monge JJ. Immunostimulants for preventing respiratory tract infection in children. Cochrane Database Syst Rev. 2006:Cd004974.

Demirkol M, Giżewska M, Giovannini M, Walter J. Follow up of phenylketonuria patients. Mol Genet Metab 2011;104:S31-39.

di Giuseppe R, de Lorgeril M, Salen P, Laporte F, Di Castelnuovo A, Krogh V, Siani A, Arnout J, Cappuccio FP, van Dongen M et al. Alcohol consumption and n-3 polyun-

saturated fatty acids in healthy men and women from 3 European populations. Am J Clin Nutr 2009;89:354–62.

Douglas RM, Hemila H, Chalker E, Treacy B. Vitamin C for preventing and treating the common cold. Cochrane Database Syst Rev. 2007:Cd000980.

Duggan C, Penny ME, Hibberd P, Gil A, Huapaya A, Cooper A, et al. Oligofructose-supplemented infant cereal: 2 randomized, blinded, community-based trials in Peruvian infants. Am J Clin Nutr. 2003;77:937-42.

Duggan C, Penny ME, Hibberd P, Gil Al, Huapaya A, Coletta F, et al. The effects of infant cereal supplemented with oligofructose in Peruvian infants: a randomized, double-blind trial. JPEN Journal of parenteral and enteral nutrition 2002.

Duijts L, Ramadhani MK, Moll HA. Breastfeeding protects against infectious diseases during infancy in industrialized countries. A systematic review. Maternal & child nutrition. 2009;5:199-210.

Eisses A. First results of a randomised controlled double blind European multi-centre study with an infant formula supplemented with immunoactive prebiotics. Part I: Effect on frequency of febrile episodes in healthy infants in the first year of life. World Congress of Pediatric Gastroenterology, Hepatology and Nutrition 2008.

Elizondo-Montemayor L, Serrano-González M, Ugalde-Casas PA, Cuello-García C, Borbolla-Escoboza JR. Plasma phospholipid fatty acids in obese male and female mexican children. Ann Nutr Metab 2010;57:234–41.

Extier A, Langelier B, Perruchot MH, Guesnet P, Van Veldhoven PP, Lavialle M, Alessandri JM. Gender affects liver desaturase expression in a rat model of n-3 fatty acid repletion. J Nutr Biochem 2010;21:180–7.

Fanaro S, Boehm G, Garssen J, Knol J, Mosca F, Stahl B, et al. Galacto-oligosaccharides and long-chain fructo-oligosaccharides as prebiotics in infant formulas: a review. Acta Paediatr Suppl. 2005;94:22-6.

Fekete K, Decsi T. Long-chain polyunsaturated fatty acids in inborn errors of metabolism. Nutrients 2010;2:965-74.

Fendrick AM, Saint S, Brook I, Jacobs MR, Pelton S, Sethi S. Diagnosis and treatment of upper respiratory tract infections in the primary care setting. Clin Ther. 2001;23:1683-706.

Fernandez-Real JM, Vayreda M, Casamitjana R, Gonzalez-Huix F, Ricart W. Circulating granulocyte-macrophage colony-stimulating factor and serum fatty acid composition in men and women. Metabolism 2001;50:1479–83.

Fleith M, Clandinin MT. Dietary PUFA for preterm and term infants: review of clinical studies. Crit Rev Food Sci Nutr 2005;45:205-29.

Galli C, Agostoni C, Mosconi C, Riva E, Salari PC, Giovannini M. Reduced plasma C-20 and C-22 polyunsaturated fatty acids in children with phenylketonuria during dietary intervention. J Pediatr 1991;119:562-7.

Geppert J, Min Y, Neville M, Lowy C, Ghebremeskel K. Gender-specific fatty acid profiles in platelet phosphatidyl-choline and -ethanolamine. Prostaglandins Leukot Essent Fatty Acids 2010;82:51–6.

Gibson GR, Probert HM, Loo JV, Rastall RA, Roberfroid MB. Dietary modulation of the human colonic microbiota: updating the concept of prebiotics. Nutrition research reviews. 2004;17:259-75.

Gibson GR, Roberfroid MB. Dietary modulation of the human colonic microbiota: introducing the concept of prebiotics. J Nutr. 1995;125:1401-12.

Giltay EJ, Gooren LJ, Toorians AW, Katan MB, Zock PL. Docosahexaenoic acid concentrations are higher in women than in men because of estrogenic effects. Am J Clin Nutr 2004;80:1167–74.

Giovannini M, Verduci E, Radaelli G et al. Long-chain polyunsaturated fatty acids profile in plasma phospholipids of hyperphenylalaninemic children on unrestricted diet. Prostaglandins Leukot Essent Fatty Acids 2011;84:39-42.

Giovannini M, Verduci E, Salvatici E, Paci S, Riva E. Phenylketonuria: nutritional advances and challenges. Nutr Metab (Lond) 2012;9:7.

Glatz JF, Soffers AE, Katan MB. Fatty acid composition of serum cholesteryl esters and erythrocyte membranes as indicators of linoleic acid intake in man. Am J Clin Nutr. 1989;49:269–76.

Glew RH, Casados JK, Huang YS, Chuang LT, VanderJagt DJ. The fatty acid composition of the serum phospholipids of children with sickle cell disease in Nigeria. Prostaglandins Leukot Essent Fatty Acids 2002;67:217–22.

Glew RH, Chuang LT, Berry T, Okolie H, Crossey MJ, VanderJagt DJ. Lipid profiles and trans fatty acids in serum phospholipids of semi-nomadic fulani in Northern Nigeria. J Health Popul Nutr 2010;28:159–66.

Goldenberg JZ, Ma SS, Saxton JD, Martzen MR, Vandvik PO, Thorlund K, et al. Probiotics for the prevention of Clostridium difficile-associated diarrhea in adults and children. Cochrane Database Syst Rev. 2013;5:Cd006095.

Gruber C, van Stuijvenberg M, Mosca F, Moro G, Chirico G, Braegger CP, et al. Reduced occurrence of early atopic dermatitis because of immunoactive prebiotics among low-atopy-risk infants. J Allergy Clin Immunol. 2010;126:791-7.

Gulani A, Sachdev HS. Zinc supplements for preventing otitis media. Cochrane Database Syst Rev. 2012;4:Cd006639.

Guyatt GH, Oxman AD, Kunz R, Atkins D, Brozek J, Vist G, Alderson P, Glasziou P, Falck-Ytter Y, Schünemann HJ. GRADE guidelines: 2. Framing the question and deciding on important outcomes. J Clin Epidemiol. 2011 Apr;64(4):395-400.

Guyatt GH, Oxman AD, Vist G, Kunz R, Brozek J, Alonso-Coello P, Montori V, Akl EA, Djulbegovic B, Falck-Ytter Y, Norris SL, Williams JW Jr, Atkins D, Meerpohl J, Schünemann HJ. GRADE guidelines: 4. Rating the quality of evidence--study limitations (risk of bias). J Clin Epidemiol. 2011 Apr;64(4):407-15.

Gwaltney JM. Clinical significance and pathogenesis of viral respiratory infections. Am J Med. 2002;112 Suppl 6A:13s-18s.

Haarman M, Knol J. Quantitative real-time PCR assays to identify and quantify fecal Bifidobacterium species in infants receiving a prebiotic infant formula. Appl Environ Microbiol. 2005;71:2318-24.

Hagenfeldt L, Hagenfeldt K, Wennmalm A. Turnover of plasma free arachidonic and oleic acids in men and women. Horm Metab Res 1975;7:467–71.

Hao Q, Lu Z, Dong BR, Huang CQ, Wu T. Probiotics for preventing acute upper respiratory tract infections. Cochrane Database Syst Rev. 2011:Cd006895.

Harnack K, Andersen G, Somoza V. Quantitation of alpha-linolenic acid elongation to eicosapentaenoic and docosahexaenoic acid as affected by the ratio of n6/n3 fatty acids. Nutr Metab 2009;6:8.

Hemila H, Louhiala P. Vitamin C for preventing and treating pneumonia. Cochrane Database Syst Rev. 2007:Cd005532.

Higgins JP, Thompson SG. Quantifying heterogeneity in a meta-analysis. Stat Med. 2002 Jun 15;21(11):1539-58.

Higgins JPT, Altman DG, Sterne JAC (editors). Chapter 8: Assessing risk of bias in included studies. In: Higgins JPT, Green S (editors). Cochrane Handbook for Systematic

Reviews of Interventions Version 5.1.0 (updated March 2011). The Cochrane Collaboration, 2011. Available from www.cochrane-handbook.org.

Higgins JPT, Deeks JJ (editors). Chapter 7: Selecting studies and collecting data. In: Higgins JPT, Green S (editors), Cochrane Handbook for Systematic Reviews of Interventions Version 5.1.0 (updated March 2011). The Cochrane Collaboration, 2011. Available from www.cochrane-handbook.org.

Higgins JPT, Green S (editors). Cochrane Handbook for Systematic Reviews of Interventions Version 5.1.0 [updated March 2011]. The Cochrane Collaboration, 2011. Available from www.cochrane-handbook.org.

Hirai K, Horiuchi R, Ohno Y, Higuchi H, Asano Y. Lower eicosapentaenoic acid and higher arachidonic acid levels in sera of young adults in the Netherlands than in Japan. Environ Health Prev Med 2000;5:60–5.

Hirai K, Kozuki M, Miyanaga K, Miyagawa F, Takezoe R, Hasegawa M, Mori M. Lower levels of eicosapentaenoic acid and the ratio of docosahexaenoic acid to arachidonic acid in sera of patients with multi-infarct dementia. J Clin Biochem Nutr 2005;36:83–9.

Hodge AM, Simpson JA, Gibson RA, Sinclair AJ, Makrides M, O'Dea K, English DR, Giles GG. Plasma phospholipid fatty acid composition as a biomarker of habitual dietary fat intake in an ethnically diverse cohort. Nutr Metab Cardiovasc Dis 2007;17:415–26.

Hodson L, Skeaff CM, Fielding BA. Fatty acid composition of adipose tissue and blood in humans and its use as a biomarker of dietary intake. Prog Lipid Res 2008;47:348–80.

Hooper L, Ashton K, Harvey LJ, Decsi T, Fairweather-Tait SJ. Assessing potential biomarkers of micronutrient status by using a systematic review methodology: methods. Am J Clin Nutr 2009;89(suppl):1953S–9S.

Infante JP, Huszagh VA. Impaired arachidonic (20:4n-6) and docosahexaenoic (22:6n-3) acid synthesis by phenylalanine metabolites as etiological factors in the neuropathology of phenylketonuria. Mol Genet Metab 2001;72(3):185-98.

Innis SM, Kuhnlein HV, Kinloch D. The composition of red cell membrane phospholipids in Canadian Inuit consuming a diet high in marine mammals. Lipids 1988;23:1064–8.

Iwamoto M, Imaizumi K, Sato M, Hirooka Y, Sakai K, Takeshita A, Kono M. Serum lipid profiles in Japanese women and men during consumption of walnuts. Eur J Clin Nutr 2002;56:629–37.

Jagannathan SN. Fatty acid composition of adipose tissue in Indian adults: sex difference and influence of pregnancy. Ind J Biochem 1969;6:222–5.

Jeurink PV, van Esch BC, Rijnierse A, Garssen J, Knippels LM. Mechanisms underlying immune effects of dietary oligosaccharides. Am J Clin Nutr. 2013;98:572S-7S.

Johnston BC, Goldenberg JZ, Vandvik PO, Sun X, Guyatt GH. Probiotics for the prevention of pediatric antibiotic-associated diarrhea. Cochrane Database Syst Rev. 2011:Cd004827.

Jolliffe DA, Griffiths CJ, Martineau AR. Vitamin D in the prevention of acute respiratory infection: systematic review of clinical studies. The Journal of steroid biochemistry and molecular biology. 2013;136:321-9.

Kale A, Joshi S, Naphade N, Sapkale S, Raju MS, Pillai A, Nasrallah H, Mahadik SP. Opposite changes in predominantly docosahexaenoic acid (DHA) in cerebrospinal fluid and red blood cells from never-medicated first-episode psychotic patients. Schiz Res 2008;98:295–301.

Karlsson M, Marild S, Brandberg J, Lonn L, Friberg P, Strandvik B. Serum phospholipid fatty acids, adipose tissue, and metabolic markers in obese adolescents. Obesity 2006;14:1931–9.

Katan MB, van Birgelen A, Deslypere JP, Penders M, van Staveren WA. Biological markers of dietary intake, with emphasis on fatty acids. Ann Nutr Metab 1991;35:249–52.

Kieu NT, Yasugi E, Hung NT, Kido T, Kondo K, Yamamoto S, Chuyen NV, Oshima M. Serum fatty acids, lipoprotein (a) and apolipoprotein profiles of middle-aged men and women in South Vietnam. Asia Pac J Clin Nutr. 2002;11:112–6.

Koletzko B, Beblo S, Demmelmair H, Hanebutt FL. Omega-3 LC-PUFA supply and neurological outcomes in children with phenylketonuria (PKU). J Pediatr Gastroenterol Nutr 2009;48:S2-7.

Koletzko B, Sauerwald T, Demmelmair H et al. Dietary long-chain polyunsaturated fatty acid supplementation in infants with phenylketonuria: a randomized controlled trial. J Inherit Metab Dis 2007;30:326-32.

Kovacs Z, Benjamins E, Grau K, Ur Rehman A, Ebrahimi M, Czermak P. Recent Developments in Manufacturing Oligosaccharides with Prebiotic Functions. Adv Biochem Eng Biotechnol. 2013.

Kuriki K, Nagaya T, Tokudome Y, Imaeda N, Fujiwara N, Sato J, Goto C, Ikeda M, Maki S, Tajima K et al. Plasma Concentrations of (n-3) Highly Unsaturated Fatty Acids Are Good Biomarkers of Relative Dietary Fatty Acid Intakes: A Cross-Sectional Study. J Nutr 2003;133:3643–50.

Lage S, Bueno M, Andrade F et al. Fatty acid profile in patients with phenylketonuria and its relationship with bone mineral density. J Inherit Metab Dis 2010; [Epub ahead of print]

Lamberti LM, Fischer Walker CL, Noiman A, Victora C, Black RE. Breastfeeding and the risk for diarrhea morbidity and mortality. BMC Public Health. 2011;11 Suppl 3:S15.

Lassi ZS, Haider BA, Bhutta ZA. Zinc supplementation for the prevention of pneumonia in children aged 2 months to 59 months. Cochrane Database Syst Rev. 2010:Cd005978.

Lee HY, Woo J, Chen ZY, Leung SF, Peng XH. Serum fatty acid, lipid profile and dietary intake of Hong Kong Chinese omnivores and vegetarians. Eur J Clin Nutr 2000;54:768–73.

Lefebvre C, Manheimer E, Glanville J. Chapter 6: Searching for studies. In: Higgins JPT, Green S (editors). Cochrane Handbook for Systematic Reviews of Interventions Version 5.1.0 (updated March 2011). The Cochrane Collaboration, 2011. Available from www.cochrane-handbook.org.

Lefebvre C, Manheimer E, Glanville J. Chapter 6: Searching for studies. In: Higgins JPT GS, editor. Cochrane Handbook for Systematic Reviews of Interventions Version 510 (updated March 2011). www.cochrane-handbook.org: The Cochrane Collaboration, 2011; 2011.

Lemaitre RN, Siscovick DS, Berry EM, Kark JD, Friedlander Y. Familial aggregation of red blood cell membrane fatty acid composition: the Kibbutzim Family Study. Metabolism 2008;57:662–8.

Lewis S, Clarke M. Forest plots: trying to see the wood and the trees. BMJ. 2001 Jun 16;322(7300):1479-80.

Liberati A, Altman D G, Tetzlaff J. et al. The PRISMA statement for reporting systematic reviews and meta-analyses of studies that evaluate healthcare interventions: explanation and elaboration. BMJ. 2009; 339 b2700

Lichtenstein AH, Yetley EA, Lau J. Application of systematic review methodology to the field of nutrition. J Nutr. 2008 Dec;138(12):2297-306. doi: 10.3945/jn.108.097154.

Lucas M, Dewailly E, Blanchet C, Gingras S, Holub BJ. Plasma omega-3 and psychological distress among Nunavik Inuit (Canada). Psych Res 2009;167:266–78.

Ma J, Folsom AR, Shahar E, Eckfeldt JH. Plasma fatty acid composition as an indicator of habitual dietary fat intake in middle-aged adults. Am J Clin Nutr. 1995;62:564–71.

MacDonald A, Rocha JC, van Rijn M, Feillet F. Nutrition in phenylketonuria. Mol Genet Metab 2011;104:S10-S18.

Mamalakis G, Kafatos A, Tornaritis M, Alevizos B. Anxiety and adipose essential fatty acid precursors for prostaglandin E1 and E2. J Am Coll Nutr 1998;17:239–43.

Mamalakis G, Kalogeropoulos N, Andrikopoulos N, Hatzis C, Kromhout D, Moschandreas J, Kafatos A. Depression and long chain n-3 fatty acids in adipose tissue in adults from Crete. Eur J Clin Nutr 2006;60:882–8.

McNamara RK. Jandacek R, Rider T, Tso P, Dwivedi Y, Pandey GN. Selective deficits in erythrocyte docosahexaenoic acid composition in adult patients with bipolar disorder and major depressive disorder. J Affect Disord 2010;126:303–11.

Melchert HU, Limsathayourat N, Mihajlovic H, Eichberg J, Thefeld W, Rottka H. Fatty acid patterns in triglycerides, diglycerides, free fatty acids, cholesteryl esters and phosphatidylcholine in serum from vegetarians and non-vegetarians. Atherosclerosis 1987;65:159–66.

Metherel AH, Armstrong JM, Patterson AC, Stark KD. Assessment of blood measures of n-3 polyunsaturated fatty acids with acute fish oil supplementation and washout in men and women. Prostaglandins Leukotr Essent Fatty Acids 2009;81:23–9.

Meyer BJ. Are we consuming enough long chain omega-3 polyunsaturated fatty acids for optimal health? Prostaglandins Leukot Essent Fatty Acids 2011;85:275-80

Mitchell EA, Lewis S, Cutler DR. Essential fatty acids and maladjusted behaviour in children. Prostaglandins Leukot Med 1983;12:281–7.

Moher D, Liberati A, Tetzlaff J, Altman DG; PRISMA Group. Preferred reporting items for systematic reviews and meta-analyses: the PRISMA statement. J Clin Epidemiol. 2009 Oct;62(10):1006-12.

Moro G, Arslanoglu S, Stahl B, Jelinek J, Wahn U, Boehm G. A mixture of prebiotic oligosaccharides reduces the incidence of atopic dermatitis during the first six months of age. Arch Dis Child. 2006;91:814-9.

Moro G, Minoli I, Mosca M, Fanaro S, Jelinek J, Stahl B, et al. Dosage-related bifidogenic effects of galacto- and fructooligosaccharides in formula-fed term infants. J Pediatr Gastroenterol Nutr. 2002;34:291-5.

Moseley K, Koch R, Moser AB. Lipid status and long-chain polyunsaturated fatty acid concentrations in adults and adolescents with phenylketonuria on phenylalanine-restricted diet. J Inherit Metab Dis 2002;25:56-64.

Moyer A (ed.). Evidence Based Pediatrics and Child Health. BMJ Books 2004, ISBN 0727917463.

Nakamura T, Takebe K, Tando Y, Arai Y, Yamada N, Ishii M, Kikuchi H, Machida K, Imamura K, Terada A. Serum fatty acid composition in normal japanese and its relationship with dietary fish and vegetable oil contents and blood lipid levels. Ann Nutr Metab1995;39:261–70.

Nikkari T, Luukkainen P, Pietinen P, Puska P. Fatty acid composition of serum lipid fractions in relation to gender and quality of dietary fat. Ann Med 1995;27:491–8.

Osborn DA, Sinn JK. Prebiotics in infants for prevention of allergy. Cochrane Database Syst Rev. 2013;3:CD006474.

Pawlosky R, Hibbeln J, Lin Y, Salem N Jr. N-3 fatty acid metabolism in women. Br J Nutr. 2003;90:993–5.

Plakké T, Berkel J, Beynen AC, Hermus RJ, Katan MB. Relationship between the fatty acid composition of the diet and that of the subcutaneous adipose tissue in individual human subjects. Hum Nutr Appl Nutr 1983;37:365–72.

Rao S, Srinivasjois R, Patole S. Prebiotic supplementation in full-term neonates: a systematic review of randomized controlled trials. Arch Pediatr Adolesc Med. 2009;163:755-64.

Ribeiro TCM, Costa-Ribeiro H, Jr., Almeida PS, Pontes MV, Leite MEQ, Filadelfo LR, et al. Stool pattern changes in toddlers consuming a follow-on formula supplemented with polydextrose and galactooligosaccharides. J Pediatr Gastroenterol Nutr. 2012;54:288-90.

Roberfroid M, Gibson GR, Hoyles L, McCartney AL, Rastall R, Rowland I, et al. Prebiotic effects: metabolic and health benefits. Br J Nutr. 2010;104 Suppl 2:S1-63.

Saadatian-Elahi M, Slimani N, Chajes V, Jenab M, Goudable J, Biessy C, Ferrari P, Byrnes G, Autier P, Peeters PH et al. Plasma phospholipid fatty acid profiles and their association with food intakes: results from a cross-sectional study within the European Prospective Investigation into Cancer and Nutrition. Am J Clin Nutr 2009;89:331–46.

Saavedra J, Tschernia A, Moore N, Abi-Hanna A, Coletta F, Emenhiser C, et al. Gastro-intestinal function in infants consuming a weaning food supplemented with oligo-fructose, a prebiotic. J Pediatr Gastroenterol Nutr 1999.

Saavedra JM, Tschernia A. Human studies with probiotics and prebiotics: clinical implications. The British journal of nutrition. 2002;87 Suppl 2:S241-6.

Salone LR, Vann WF, Jr., Dee DL. Breastfeeding: an overview of oral and general health benefits. Journal of the American Dental Association (1939). 2013;144:143-51.

Sanjurjo P, Perteagudo L, Rodríguez Soriano J, Vilaseca A, Campistol J. Polyunsaturated fatty acid status in patients with phenylketonuria. J Inherit Metab Dis 1994;17:704-9.

Schmelzle H, Wirth S, Skopnik H, Radke M, Knol J, Bockler HM, et al. Randomized double-blind study of the nutritional efficacy and bifidogenicity of a new infant formula containing partially hydrolyzed protein, a high beta-palmitic acid level, and non-digestible oligosaccharides. J Pediatr Gastroenterol Nutr. 2003;36:343-51.

Seifert S, Watzl B. Inulin and oligofructose: review of experimental data on immune modulation. J Nutr. 2007;137:2563S-67S.

Sfar S, Laporte F, Braham H, Jawed A, Amor S, Kerkeni A. Influence of dietary habits, age and gender on plasma fatty acids levels in a population of healthy Tunisian subjects. Exp Geront 2010;45:719–25.

Smit EN, Fokkema MR, Boersma ER, Muskiet FA. Higher erythrocyte 22:6n-3 and 22:5n-6, and lower 22:5n-3 suggest higher Delta-4-desaturation capacity in women of childbearing age. Br J Nutr 2003;89:739–40.

Srinivasjois R, Rao S, Patole S. Prebiotic supplementation in preterm neonates: Updated systematic review and meta-analysis of randomised controlled trials. Clin Nutr. 2013.

Srinivasjois R, Rao S, Patole S. Prebiotic supplementation of formula in preterm neonates: a systematic review and meta-analysis of randomised controlled trials. Clin Nutr. 2009;28:237-42.

Stuijvenberg M, Eisses AM, Grüber C, Mosca F, Arslanoglu S, Chirico G, et al. Do prebiotics reduce the number of fever episodes in healthy children in their first year of life: a randomised controlled trial. The British journal of nutrition 2011 doi: 10.1017/S0007114511004053.

Sutherland WH, Shilton ME, Nye ER, Gillies ME, Bakani I, Robertson MC. Urban/rural differences in red blood cell fatty acid composition, plasma lipids and diet in Melanesian Fijians. Eur J Clin Nutr 1995;49:233–41.

Szajewska H. Importance of Systematic Reviews and Meta-Analyses in Pediatric Nutrition. in: Szajewska H, Shamir R (eds): Evidence-Based Research in Pediatric Nutrition. World Rev Nutr Diet. Basel, Karger, 2013, vol 108, pp 1-10 (DOI:10.1159/000351479)

Takita T, Nakamura K, Kimira M, Yamada N, Kobayashi Y, Innami S. Serum fatty acid compositions and lipid concentrations and their correlations. J Clin Biochem Nutr 1996;20:149–59.

Tavendale R, Lee AJ, Smith WC, Tunstall-Pedoe H. Adipose tissue fatty acids in Scottish men and women: results from the Scottish Heart Health Study. Atherosclerosis 1992;94:161–9.

Thomas DW, Greer FR, American Academy of Pediatrics Committee on N, American Academy of Pediatrics Section on Gastroenterology H, Nutrition. Probiotics and prebiotics in pediatrics. Pediatrics. 2010;126:1217-31.

Tjønneland A, Overvad K, Thorling E, Ewertz M. Adipose tissue fatty acids as biomarkers of dietary exposure in Danish men and women. Am J Clin Nutr 1993;57:629–33.

Tschernia A, Moore N, Abi-Hanna A, Yolken R, Coletta F, Emenhiser C, et al. Effects of long-term consumption of a weaning food supplemented with oligofructose, a prebiotic, on general infant health status. J Pediatr Gastroenterol Nutr 1999.

Umemura U, Ishimori M, Kobayashi T, Tamura Y, Koike KA, Shimamoto T. Possible effects of diets on serum lipids, fatty acids and blood pressure levels in male and female Japanese university students. Environ Health Prev Med 2005;10:42–7.

Vallés J, Aznar J, Santos MT. Composition of platelet fatty acids and their modulation by plasma fatty acids in humans: effect of age and sex. Atherosclerosis 1988;71:215–25.

van Gool CJ, van Houwelingen AC, Hornstra G. The essential fatty acid status in phenylketonuria patients under treatment. J Nutr Biochem 2000;11:543-7.

Van Zutphen K, Packman W, Sporri L, Needham M, Morgan C, Weisiger K, Packman S. Executive functioning in children and adolescents with phenylketonuria. Clin Genet 2007;72(1):13-8.

Vilaseca MA, Lambruschini N, Gómez-López L et al. Long-chain polyunsaturated fatty acid status in phenylketonuric patients treated with tetrahydrobiopterin. Clin Biochem 2010;43:411-5.

Walker CL, Rudan I, Liu L, Nair H, Theodoratou E, Bhutta ZA, et al. Global burden of childhood pneumonia and diarrhoea. Lancet. 2013;381:1405-16.

Warensjö E, Ohrvall M, Vessby B. Fatty acid composition and estimated desaturase activities are associated with obesity and lifestyle variables in men and women. Nutr Metab Cardiovasc Dis 2006;16:128–36.

Wennberg M, Bergdahl IA, Hallmans G, Norberg M, Lundh T, Skerfving S, Strömberg U, Vessby B, Jansson JH. Fish consumption and myocardial infarction: a second prospective biomarker study from northern Sweden. Am J Clin Nutr 2011;93:27–36.

Wennberg M, Bergdahl IA, Stegmayr B, Hallmans G, Lundh T, Skerfving S, Strömberg U, Vessby B, Jansson JH. Fish intake, mercury, long-chain n-3 polyunsaturated fatty acids and risk of stroke in northern Sweden. Brit J Nutr 2007;98:1038–45.

Whelan K. Probiotics and prebiotics in the management of irritable bowel syndrome: a review of recent clinical trials and systematic reviews. Curr Opin Clin Nutr Metab Care. 2011;14:581-7.

Yamada T, Strong JP, Ishii T, Ueno T, Koyama M, Wagayama H, Shimizu A, Sakai T, Malcom GT, Guzman MA. Atherosclerosis and omega-3 fatty acids in the populations of a fishing village and a farming village in Japan. Atherosclerosis 2000;153:469–81.

Yeh LL, Kuller LH, Bunker CH, Ukoli FA, Huston SL, Terrell DF. The role of socioeconomic status and serum fatty acids in the relationship between intake of animal foods and cardiovascular risk factors. Ann Epidemiol 1996;6:290–8.

Yi SH, Kable JA, Evatt ML, Singh RH. A cross-sectional study of docosahexaenoic acid status and cognitive outcomes in females of reproductive age with phenylketonuria. J Inherit Metab Dis 2011;34:455-63.

Yi SH, Kable JA, Evatt ML, Singh RH. A randomized, placebo-controlled, double-blind trial of supplemental docosahexaenoic acid on cognitive processing speed and executive function in females of reproductive age with phenylketonuria: A pilot study. Prostaglandins Leukot Essent Fatty Acids 2011;85:317-27.

8. List of publications

Publications on which the thesis was based

- Lohner S, Fekete K, Marosvölgyi T, Decsi T: Gender differences in long-chain polyunsaturated fatty acid status: systematic review of 51 publications. Ann Nutr Metab. 2013;62(2):98-112 (IF₂₀₁₂: 1.661)
- Lohner S, Fekete K, Decsi T. Lower n-3 long-chain polyunsaturated fatty acid values in patients with phenylketonuria: a systematic review and meta-analysis. Nutr Res. 2013 Jul;33(7):513-20. (IF₂₀₁₂: 2.142)
- Lohner S, Küllenberg D, Antes G, Decsi T, Meerpohl JJ. Prebiotics in healthy infants and children for prevention of acute infectious diseases: a systematic review and meta-analysis. Nutr Rev. 2014. Accepted for publication (IF₂₀₁₂: 4.597)

Other publications

- Lohner S, Vágási J, Marosvölgyi T, Tényi T, Decsi T. Inverse association between 18-carbon trans fatty acids and intelligence quotients in smoking schizophrenia patients. Psychiatry Res. 2014 Jan 30;215(1):9-13. (IF₂₀₁₂: 2.456)
- Lohner Sz, Vágási J, Péterfia Cs, Decsi T. Halolajat tartalmazó étrendkiegészítők szerepe az atópiás betegségek kezelésében. Gyermekorvos Továbbképzés 2013; XII (5):221-223
- Płudowski P, Karczmarewicz E, Bayer M, Carter G, Chlebna-Sokół D, Czech-Kowalska J, Dębski R, Decsi T, Dobrzańska A, Franek E, Głuszko P, Grant WB, Holick MF, Yankovskaya L, Konstantynowicz J, Książyk JB, Księżopolska-Orłowska

K, Lewiński A, Litwin M, *Lohner S*, Lorenc RS, Lukaszkiewicz J, Marcinowska-Suchowierska E, Milewicz A, Misiorowski W, Nowicki M, Povoroznyuk V, Rozentryt P, Rudenka E, Shoenfeld Y, Socha P, Solnica B, Szalecki M, Tałałaj M, Varbiro S, Żmijewski MA. *Practical guidelines for the supplementation of vitamin D and the treatment of deficits in Central Europe - recommended vitamin D intakes in the general population and groups at risk of vitamin D deficiency.* Endokrynol Pol. 2013;64(4):319-27. (IF₂₀₁₂: **1.070**)

- Fekete K, Berti C, Trovato M, *Lohner S*, Dullemeijer C, Souverein OW, Cetin I, Decsi T: *Effect of folate intake on health outcomes in pregnancy: a systematic review and meta-analysis on birth weight, placental weight and length of gestation*. Nutrition Journal. 2012, 11:75 (IF₂₀₁₂: **2.648**)
- Lohner S, Fekete K, Berti C, Hermoso M, Cetin I, Koletzko B, Decsi T. Effect of folate supplementation on folate status and health outcomes in infants, children and adolescents: A systematic review. Int J Food Sci Nutr. 2012 Dec; 63(8):1014-1020 (IF₂₀₁₂: 1.257)
- Lohner Sz, Marosvölgyi T, Burus I, Schmidt J, Molnár D, Decsi T. Elhízott gyer-mekek étrendjének kiegészítése napi 1000 mg alfa-linolénsavval. Placebóval kontrollált, kettősen vak vizsgálat (Dietary supplementation of obese children with 1000 mg alpha-linolenic acid per day: a placebo-controlled double blind study, in Hungarian with English summary); Orvosi Hetilap 148(32):1499-1513, 2007.
- Lohner Sz, Marosvölgyi T, Schmidt J, Molnár D, Decsi T. Az alfa-linolénsav jelentősége a gyermekkori elhízáshoz társuló fokozott kardiovaszkuláris kockázat csökkentésében (Role of alpha-linolenic acid in amelioration of the

cardiovascular risk related to obesity, in Hungarian with English summary); Gyermekgyógyászat 57(3): 345-350, 2006.

• Marosvölgyi T, Kovács A, Lohner Sz, Funke S, Burus I, Decsi T. Az anyatej zsír-savösszetétele koraszülöttet és érett újszülöttet szülő anyákban a szoptatás első három hetében (Fatty acid composition of human milk in mothers of preterm and full-term infants in the first three weeks fo lactation, in Hungarian

with English summary); Orvosi Hetilap 147(31):1459-1463, 2006.

Szabó É, Lohner Sz, Molnár D, Decsi T. A transz izomér telítetlen zsírsavak kedvezőtlen hatásai a perinatális időszakban (Unfavorable effects of trans isomeric fatty acids in the perinatal period, in Hungarian); Gyermekorvos Továbbképzés IV(4): 48-51, 2006.

Cumulative impact factor of publications: 15.831

Cumulative impact factor of publications as first author: 12.113

Book chapters

■ Lohner S, Decsi T. Role of Long-Chain Polyunsaturated Fatty Acids in the Prevention and Treatment of Atopic Diseases. In: Polyunsaturated Fatty Acids: Sources, Antioxidant Properties and Health Benefits (edited by: Angel Catalá). NOVA Publishers. 2013. Chapter 11, pp. 1-24. (ISBN 978-1-62948-151-7).

Lohner Sz, Vágási J, Decsi T. Long-chain polyunsaturated fatty acid supplementation in the treatment of children with atopic dermatitis. In: 10. Országos Interdiszciplináris Grastyán Konferencia előadásai (edited by: Szamonek V). 2012, pp. 207-212. (ISBN 978-963-642-471-8).

- Vágási J, Lohner Sz, Marosvölgyi T, Tényi T, Decsi T. A többszörösen telítetlen zsírsavakkal való ellátottság vizsgálata szkizofrén betegekben (Long-chain polyunsaturated fatty acid status of patients with schizophrenia, in Hungarian). In: 10. Országos Interdiszciplináris Grastyán Konferencia előadásai (edited by: Szamonek V). 2012, pp. 382-389. (ISBN 978-963-642-471-8).
- Lohner Sz, Fekete K, Decsi T. A folsav szupplementáció gyermekgyógyászati vonatkozásai: eredmények és további lehetőségek (Paediatric aspects of folic acid supplementation: results and further possibilities, in Hungarian with English summary). In: Interdiszciplináris Doktorandusz Konferencia 2012 Konferenciakötet (edited by: Sipos N, Gunszt D). pp. 261-269. (ISBN 978-963-642-484-8).

Oral presentations

- **Lohner Szimonetta:** Veleszületett anyagcsere-betegségben szenvedő gyermekek étrendjének kiegészítése. Magyar Gyermekorvosok Társasága 2014. évi Jubileumi Nagygyűlése, 2014 május, Budapest
- Szili Nóra, Soltész Dorottya, Jakobik Viktória, Mihályi Krisztina, *Lohner Szimonetta*, Decsi Tamás: *Óvodáskorú gyermekek étkezésének energia- és tápanyag tartalma a táplálkozási ajánlások tükrében*. Magyar Gyermekorvosok Társasága 2014. évi Jubileumi Nagygyűlése, 2014 május, Budapest
- Krisztina Mihályi, Eszter Györei, Éva Szabó, Szimonetta Lohner, Tamás Marosvölgyi,
 Tamás Decsi: Changes of the fatty acid composition of breast milk during the lactation
 47th Annual Meeting of the European Society for Paediatric Gastroenterology,
 Hepatology and Nutrition, 2013 May, London
- **Lohner Szimonetta**, Fekete Katalin, Decsi Tamás: *A fehérjeszegény diétát tartó* fenilketonúriás betegek n-3 többszörösen telítetlen zsírsavakkal való ellátottsága nem

- megfelelő. XI. Országos Interdiszciplináris Grastyán Konferencia, 2013. április, Pécs (*A legközérthetőbb tudományos előadás díja*)
- Péterfia Csaba, Lohner Szimonetta, Decsi Tamás: Az eikozanoidok szerepe az asztmában zajló légúti gyulladás patogenezisében. A Magyar Tüdőgyógyász Társaság
 57. Nagygyűlés és 100 éves Centenáriumi Emlékülése, 2012. június, Budapest
- *Szimonetta Lohner*, Judit Vágási, Tamás Decsi: *Dietary omega-3 or omega-6 fatty acids for prevetion and treatment of atopy and asthma.* 4th International Interdisciplinary Grastyán Conference, 2012 April, Pécs
- Vágási Judit, Lohner Szimonetta, Marosvölgyi Tamás, Tényi Tamás, Decsi Tamás: A többszörösen telítetlen zsírsavakkal való ellátottság vizsgálata szkizofrén betegekben.
 X. Országos Interdiszciplináris Grastyán Konferencia, 2012. április, Pécs
- **Lohner Szimonetta**, Decsi Tamás: *Vitamin D status in Hungary –novel evidence for supplementation*. 1st International Dotoral Workshop on Natural Sciences, University of Pécs, 2012
- *Szimonetta Lohner*, Tamás Decsi: *Vitamin D deficiency an overview of Hungarian epidemiologic data.* Vitamin D Minimum, maximum, optimum. 2012. October,
 Warsaw
- *Lohner Szimonetta*, Fekete Katalin, Marosvölgyi Tamás, Decsi Tamás: *Nemi eltérések* a plazma- és vörösvértest membrán lipidek zsírsavösszetételében. MGYT-MGT Gyermekgasztroenterológiai Szekció XXVIII. Tudományos Ülése, 2011. szeptember, Hévíz
- *Lohner Szimonetta*, Fekete Katalin, Marosvölgyi Tamás, Decsi Tamás: *Van-e nemi különbség a plazmalipidek zsírsavösszetételében? -szisztematikus irodalmi áttekintés.*MGYT Éves Nagygyűlése, 2011. szeptember, Pécs
- **Lohner Szimonetta**, Decsi Tamás: *Gamma-linolénsavval kiegészített étrend alkalma*zása atópiás dermatitises gyermekek bőrtüneteinek befolyásolására. A Magyar Táplálkozástudományi Társaság XXXIII. Vándorgyűlése, 2008. október, Pécs

- **Lohner Szimonetta**, Szinku Ilona: *A keresztvédelem szerepe a rotavírus elleni vakcinációban*. A Pécsi Paediater Érdekvédelmi Egyesület továbbképző délutánja, 2008.február, Pécs
- **Lohner Szimonetta**, Decsi Tamás: *Omega-6 zsírsavszupplementáció atópiás* dermatitises gyermekekben szisztémás irodalmi áttekintés. Fiatal Gyermekgyógyászok Konferenciája, 2008. március, Pécs
- **Lohner Szimonetta**, Mosdósi Bernadett, Decsi Tamás: *A humorális immunitás ritka rendellenesége periódikusan visszatérő láz hátterében*. Fiatal Gyermekgyógyászok Konferenciája, 2007. március, Miskolc
- *Lohner Szimonetta*, Mosdósi Bernadett, Decsi Tamás: *Periódikus láz*. A Pécsi Paediater Érdekvédelmi Egyesület és a BM Kórház, Kerpel-Fronius Ödön Gyermekegészségügyi Központ továbbképző délutánja, 2007. február, *Pécs*
- **Lohner Szimonetta**, Hádzsiev Kinga, Decsi Tamás: *Hosszú távú biszfoszfonát kezelés* osteogenesis imperfectában a szubjektív panaszok és objektív tünetek csökkentésére. Fiatal Gyermekgyógyászok Konferenciája, 2006. április, Debrecen
- *Lohner Szimonetta*, Marosvölgyi Tamás, Schmidt János, Molnár Dénes, Decsi Tamás: *Elhízott gyermekek zsírsavellátottságának befolyásolása speciális étrenddel.* MGYT-MGT Gyermekgasztroenterológiai Szekció XXII. Tudományos Ülése, 2005. október, Eger

Poster presentations

- *Szimonetta Lohner*, Katalin Fekete, Tamás Marosvölgyi, Tamás Decsi: *Is gender a relevant modifying factor influencing the polyunsaturated fatty acid composition of serum and erythrocyte membrane lipids in all age groups?* 47th Annual Meeting of the European Society for Paediatric Gastroenterology, Hepatology and Nutrition, 2013 May, London *(ESPGHAN Young Investigators Award 2013)*